

Book Chapter

Oxidative Stress-Responsive MicroRNAs in Heart Injury

Branislav Kura¹, Barbara Szeiffova Bacova¹, Barbora Kalocayova¹, Matus Sykora^{1,2} and Jan Slezak^{1*}

¹Centre of Experimental Medicine, Institute for Heart Research, Slovak Academy of Sciences, Slovakia

²Department of Animal Physiology and Ethology, Faculty of Natural Sciences, Comenius University, Slovakia

***Corresponding Author:** Jan Slezak, Centre of Experimental Medicine, Institute for Heart Research, Slovak Academy of Sciences, 841 04 Bratislava, Slovakia

Published **June 15, 2020**

This Book Chapter is a republication of an article published by Jan Slezak, et al. at International Journal of Molecular Sciences in January 2020. (Kura, B.; Szeiffova Bacova, B.; Kalocayova, B.; Sykora, M.; Slezak, J. Oxidative Stress-Responsive MicroRNAs in Heart Injury. *Int. J. Mol. Sci.* 2020, 21, 358.)

How to cite this book chapter: Branislav Kura, Barbara Szeiffova Bacova, Barbora Kalocayova, Matus Sykora, Jan Slezak. Oxidative Stress-Responsive MicroRNAs in Heart Injury. In: Song Guo Zheng, editor. Prime Archives in Molecular Biology. Hyderabad, India: Vide Leaf. 2020.

© The Author(s) 2020. This article is distributed under the terms of the Creative Commons Attribution 4.0 International License(<http://creativecommons.org/licenses/by/4.0/>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Author Contributions: J.S. supervised the writing project of the manuscript; B.K. (Branislav Kura), B.K. (Barbora Kalocayova),

and B.S.B. prepared the manuscript and wrote the draft together; M.S. prepared the figures. All authors have read and agreed to the published version of the manuscript.

Funding: This research was funded by a grant from the Slovak Research and Development Agency of Ministry of Education, Science, Research, and Sport of the Slovak Republic (APVV-15-0376) and grants from the Scientific Grant Agency of the Ministry of Education, Science, Research, and Sport of the Slovak Republic and the Slovak Academy of Sciences (VEGA 2/0021/15, VEGA 2/0063/18, VEGA 2/0158/19, VEGA 2/0166/17, VEGA 2/0002/20) and grant from European Union Structural funds (ITMS 26230120009).

Acknowledgments: The authors thanks all of the individuals who participated in the investigations.

Conflicts of Interest: The authors declare no conflicts of interest.

Abstract

Reactive oxygen species (ROS) are important molecules in the living organisms as a part of many signaling pathways. However, if overproduced, they also play a significant role in the development of cardiovascular diseases, such as arrhythmia, cardiomyopathy, ischemia/reperfusion injury (e.g., myocardial infarction and heart transplantation), and heart failure. As a result of oxidative stress action, apoptosis, hypertrophy, and fibrosis may occur. MicroRNAs (miRNAs) represent important endogenous nucleotides that regulate many biological processes, including those involved in heart damage caused by oxidative stress. Oxidative stress can alter the expression level of many miRNAs. These changes in miRNA expression occur mainly via modulation of nuclear factor erythroid 2-related factor 2 (Nrf2), sirtuins, calcineurin/nuclear factor of activated T cell (NFAT), or nuclear factor kappa B (NF- κ B) pathways. Up until now, several circulating miRNAs have been reported to be potential biomarkers of ROS-related cardiac diseases, including myocardial infarction, hypertrophy, ischemia/reperfusion, and

heart failure, such as miRNA-499, miRNA-199, miRNA-21, miRNA-144, miRNA-208a, miRNA-34a, etc. On the other hand, a lot of studies are aimed at using miRNAs for therapeutic purposes. This review points to the need for studying the role of redox-sensitive miRNAs, to identify more effective biomarkers and develop better therapeutic targets for oxidative-stress-related heart diseases.

Keywords

Cardiovascular Diseases; Ischemia/Reperfusion Injury; miRNA; Oxidative Stress; Transplantation

Introduction

Despite advances in disease prevention, diagnosis, and treatment, cardiovascular diseases (CVDs) are still in the leading position as the cause of mortality and morbidity worldwide. It is estimated that by 2030, nearly 23.6 million people will die from CVDs, primarily from heart disease and stroke, per year [1,2]. As both reactive oxygen species (ROS) production and microRNA (miRNA) expression signature have been associated with the development of CVDs, it is important to understand the crosstalk between ROS and miRNAs [3,4].

ROS are constantly released during mitochondrial oxygen consumption for energy production. Any imbalance between ROS production and its scavenger system induces oxidative stress. Oxidative stress, a critical contributor to tissue damage, is well-known to be associated with various diseases [5]. It has been long recognized that an increase of ROS can modify the cell-signaling proteins and has functional consequences, which mediate pathological processes included in the development of CVDs related to hypoxia, cardiotoxicity, and ischemia-reperfusion [6]. It was reported that myocardial ROS levels were elevated in animal models of ischemia/reperfusion injury [7] and heart failure [8]. Recent data from T. Wongsurawat study reveal elevated level of ROS species at the proteomic and transcriptomic level in the vessel wall [9-11].

MiRNAs are integrated into a group of small, naturally occurring and noncoding RNAs (size 21–25 nucleotides), which modulate gene expression at the post-transcriptional level. MiRNAs play a role as regulators of gene expression through binding to complementary sequences on the 3'-untranslated region (3'-UTR) of their target mRNA, thus inhibiting mRNA translation or promoting mRNA degradation [12,13]. Many miRNA genes are transcribed by enzyme RNA polymerase II from intergenic, intronic, or polycistronic loci as a long primary miRNA transcript (pri-miRNA), which is then cleaved by the enzyme Drosha endoribonuclease to a 70-nt-long hairpin structure with 2-nt-30 overhangs (pre-miRNA). Pre-miRNA is thereafter exported to the cytoplasm and processed by a second endoribonuclease enzyme (Dicer), to form a 22-nucleotide-long miRNA:miRNA* duplex with partial complementarity. One strand of this duplex then combines with the Argonaute (AGO) protein into the RNA-induced silencing complex (RISC), while the passenger strand gets degraded. One mRNA can contain multiple binding sites for different miRNAs, thus creating a complicated network of miRNA–mRNA interactions. MiRNAs are distributed in tissue-specific patterns and are able to regulate the expression of approximately 30% of human genes [14-16].

Numerous studies have shown that miRNAs have essential roles in cardiovascular development, pathology, regeneration, and repair and could be used for the diagnosis and prevention of cardiovascular diseases, such as hypertrophy, myocardial infarction, contractility defects, arrhythmias, etc. [3,17-21]. In response to increased ROS or stress stimuli, CVDs are obviously initiated and progressed by apoptosis, autophagy, necrosis, and fibrosis, as well as proliferation and migration of cardiomyocytes and endothelial cells, cardiac fibroblasts, and vascular smooth muscle cells. It was documented that miRNAs are involved in these processes [3,18,22]. Moreover, some miRNAs have been assigned as regulators of oxidative stress in the cardiovascular system by targeting ROS generators, antioxidant signaling pathways, and selected antioxidant effectors [23].

In this review, we tried to highlight recent findings about the association of miRNAs with the development of CVDs,

including atherosclerosis, myocardial infarction, cardiac hypertrophy, or heart failure. There is evidence of interactions between cardiac miRNAs and ROS, but further studies need to be provided to reveal the molecular mechanisms of miRNAs regulating CVD diseases under ROS-related stress conditions.

Oxidative Stress and Cardiovascular System

The common pathological feature of most cardiac and vascular diseases is an imbalance of biological system in oxidation and antioxidation or between the generation and detoxification of ROS, which is generally called oxidative stress [24,25].

ROS are small reactive molecules implicated in the regulation of various cell functions and biological processes [26]. They are defined as molecules containing at least one atom of oxygen with higher reactivity than molecular oxygen, like superoxide anion (O_2^-), hydrogen peroxide (H_2O_2), hydroxyl radical (OH^\cdot), peroxyxynitrite ($ONOO^-$), hypochlorous acid ($HOCl$), and others [27]. In smaller or moderate concentrations, ROS can serve as signaling molecules, but an uncontrolled higher level of ROS leads to free-radical damage associated with the structural and functional alterations of proteins, lipids, and deoxyribonucleic acid [28,29].

The generation of ROS could be divided into two categories. Firstly, they are produced mainly by mitochondrial oxidative metabolism as a by-product or waste product, and secondly, as a cellular response to stress, xenobiotics, cytokines, and bacterial invasion, where ROS are formed intentionally as part of a signal transduction pathway [6,30]. Most relevant enzymatic sources of ROS are the nicotinamide adenine dinucleotide phosphate (NADPH) oxidases (NOXs), xanthine oxidase, uncoupled nitric oxide (NO) synthase, and mitochondria [24]. To the other incentives belong tumor necrosis factor-alpha ($TNF-\alpha$), epidermal growth factor, Interleukin-1beta ($IL-1\beta$), hypoxia, and irradiation [6].

To ensure homeostasis in these processes, proteins with an antioxidant activity that protect an aerobic organism from

increased toxicity of ROS have been found [31]. Major antioxidant proteins are superoxide dismutases (SODs), which catalyze conversion of superoxide into oxygen and hydrogen peroxide [32]. Hydrogen peroxide produced by SOD is then subsequently transformed into water and oxygen by other antioxidant proteins, glutathione peroxidases, catalases, and thioredoxins [33]. Taken together, to keep redox equilibrium is very difficult due to a variety of mechanisms, and, in the case of the antioxidant cell defense system that is suppressed by oxidative stress, many diseases can occur, especially in the cardiovascular system, where oxygen delivery to myocardium is approximately 1.6–1.8 times higher than in other tissues [34].

Increased levels of ROS are unfavorably implicated in the myocardial calcium handling, cardiac remodeling induced by hypertrophic signaling, apoptosis, and necrosis. Oxidative stress similarly has a negative effect on blood vessels, their function, angiogenesis, apoptosis, vascular tone, and genomic stability [24]. This suggests that cardiovascular risk factors with elevated ROS levels are interconnected with endothelial dysfunction. Dysregulated generation of ROS also contributes to the pathogenesis of atherosclerosis, heart failure, cardiomyopathy, and cardiac hypertrophy [35] (Figure 1).

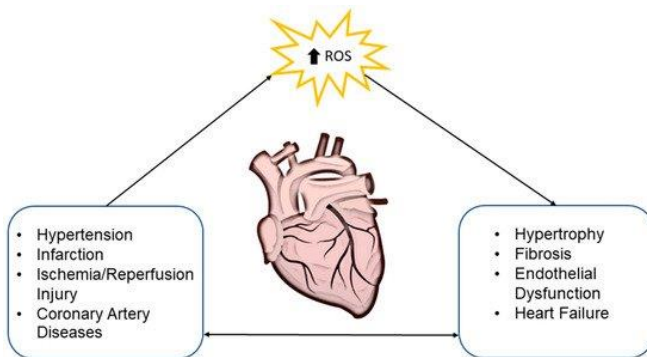


Figure 1: Impact of oxidative stress on the heart. Overproduction of reactive oxygen species (ROS) contributes to different cardiac pathologies, e.g., hypertrophy or fibrosis. These pathological changes in the heart may result in cardiovascular diseases, which may subsequently contribute to the production of ROS.

It was observed that activation of neuroendocrine pathways, namely sympathetic and the renin-angiotensin-aldosterone system in patients with failing myocardium, was associated with oxidative stress [36,37]. In experimental and human studies dealing with heart failure, the elevated activity of NOX has been consistently observed [37,38]. For example, the implication of NOX isoforms in the development of left ventricular hypertrophy (LVH) was demonstrated in NOX2 knockout mice, where infusion of Angiotensin II resulted in less incidence of LVH compared to control wild-type mice infused with the same concentration of Angiotensin II induced LVH [39]. A similar conclusion was drawn from neonatal rat ventricular cardiomyocytes where activation of NOX2 was associated with angiotensin II-induced cardiac hypertrophy [40]. Besides that, inactivation of NOX2 leads to the reduction of infarct size in a model of myocardial infarction [41]. Activation of NOX4 also has an impact on heart failure [42]. This implication was shown in experimental study with transgenic mice with cardiac-specific overexpression of NOX4, where incidence of fibrosis, apoptosis, and enlargement of cardiomyocytes were found [43]. Besides, upregulated expression of NOX4 was associated with overexpression of lysocardiolipin acyltransferase-1, which was implicated in the catalysis of cardiolipin synthesis [44]. Upregulated lysocardiolipin acyltransferase-1 by oxidative stress damages phospholipid cardiolipin, a component of the inner mitochondrial membrane leading to increased levels of O_2^- , $ONOO^-$, and NO radicals, which results in the decline of ATP production and disorganization of the dimeric ADP/ATP carrier functional capacity [45,46]. Unlike this, in mice with cardiac-specific deletion of NOX4 with applied transverse aortic constriction, the pathological changes were less pronounced [43]. Expression and activity of xanthine oxidase were also increased in patients with heart failure, whereas inhibition of this enzyme ameliorated heart contractility, as well as endothelial dysfunction [47].

Hypertension, hypercholesterolemia, hyperglycemia, and atherosclerosis are also typical with the activation of ROS enzyme sources [24]. Elevated ROS levels, activation of NOX [48], impairment of NO/cGMP signaling, and subsequently

reduced acetylcholine-mediated vasodilation in hypertension models were induced by Angiotensin II stimulation [49]. Moreover, a decline in superoxide dismutase and glutathione peroxidase activity were inversely correlated with blood pressure in untreated hypertensive patients [50]. The process of atherosclerosis is closely linked with lack of NO production or its accelerated scavenging [51]. Generated ONOO⁻ induces transformation of smooth muscle cells into foam cells, as well as release of matrix metalloproteinases, which degrade atheromatous plaque and basement membrane of the endothelial cells leading to physical disruption of the plaques [26].

Another situation of oxidative-stress-related cardiac damage is heart transplantations, where the donor heart, graft, is exposed to cold ischemia-reperfusion injury associated with increased ROS. This leads to graft dysfunction, like allograft rejection, delayed graft function, or primary nonfunction, as well as to endothelial and parenchymal cell injury [52,53]. To increase success of the heart transplantation, experimental studies have focused on the enrichment of antioxidative substrates in cardioplegia solution and pump prime solution [54-59], suggesting the importance of continuous research into antioxidants.

Oxidative Stress and MiRNA

Many previous studies have demonstrated that multiple molecular mechanisms and signaling pathways can regulate oxidative stress [60]. Growing evidence has confirmed that miRNAs can be considered as potential targets and modulators of oxidative-stress-related pathways [61]. By analysis of miRNA expression signature implicated in oxidative stress-related pathways, several miRNAs were identified and termed as oxidative stress-responsive miRNAs [62]. The increasing number of studies shows that intracellular ROS can either inhibit or induce miRNA expression level, which results in subsequent biological effects through regulation of their direct target genes (Figure 2) [63]. Among them, several pathways (Nrf2—nuclear factor erythroid 2-related factor 2, SIRT1—sirtuin 1, and NF- κ B—nuclear factor kappa B) have been the most intensively

studied in connection with oxidative stress and miRNA. These will be further described in this section.

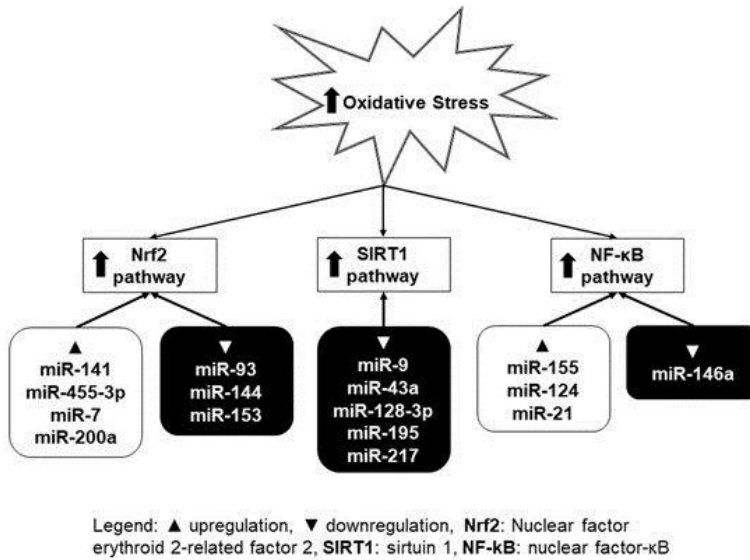


Figure 2: Selected signaling pathways (Nrf2, SIRT1, and NF-κB) influenced by miRNAs in situations with oxidative stress. ROS either inhibit or induce miRNA expression level. This leads to subsequent regulation of their target genes. Upper arrow represents increase of oxidative stress/activation of signaling pathway.

Nrf2 Pathway

Nrf2 plays a part in the cellular antioxidant defense system by upregulating the expression levels of antioxidant enzymes [64], such as glutathione S-transferase (GST), NAD(P)H:quinone oxidoreductase (NQO) 1, SOD1, and heme oxygenase (HO) 1, through binding to antioxidant response elements (AREs) in their promoters [23]. Under physiological conditions, Nrf2 is bound to its inhibitory protein, Kelch-like ECH-associated protein 1 (Keap1), which limits its transcriptional activity in the nucleus. Oxidative stress causes Nrf2 to dissociate from Keap1, which results in its binding to ARE and the transcription of downstream target genes [65].

Some miRNAs were reported to target Nrf2 directly. Zhu et al. revealed that miRNA-153 promotes oxidative stress by negatively regulating Nrf2 in an in vitro model of Parkinson's disease [66]. The study of Sangokoya et al. shows that increased expression of miRNA-144 is associated with reduced Nrf2 levels in homozygous sickle cell disease (HbSS) reticulocytes and with decreased glutathione regeneration and attenuated antioxidant capacity in HbSS erythrocytes [67]. Another study found that the downregulation of miRNA-93 elevates Nrf2 expression and alleviates reactive oxygen species and cell apoptosis in diabetic retinopathy [68].

Besides directly targeting Nrf2, miRNAs may also target its regulators. Cheng et al. demonstrated that miRNA-141 attenuates UV-induced oxidative stress via targeting Keap1 to activate Nrf2 signaling in human retinal pigment epithelium cells and retinal ganglion cells [69]. MiRNA-7 represses Keap1 expression in human neuroblastoma cells, decreases the intracellular hydroperoxide level, and increases the level of the reduced form of glutathione [70]. Zhang et al. demonstrated that miRNA-455-3p activated the Nrf2/ARE signal pathway through suppressing Keap1, thereby suppressing oxidative stress and promoting osteoblasts growth [71]. Other authors showed that miRNA-200a controls Nrf2 activation by target Keap1 in hepatic stellate cell proliferation and fibrosis [72].

SIRT1 Pathway

The increasing number of studies confirms that SIRT1 is an important component of cellular responses to oxidative stress [73,74]. SIRT1 is a target of various redox-sensitive pathways [23]. To induce an antioxidant response, activated SIRT1 deacetylates multiple targets, including endothelial nitric oxide synthase (eNOS), peroxisome proliferator-activated receptor- γ coactivator 1- α (PGC1 α), p53, Forkhead box O transcription factors (FoxO), Nrf2, and NF- κ B [65]. In the cell, the deacetylation of FoxO1 by SIRT1 increases transcriptional activity and upregulates downstream antioxidants such as SOD2 and catalase [75].

Several miRNAs have been reported to influence oxidative stress by directly targeting SIRT1. Downregulation of SIRT1 by miRNA-34a promoted vascular smooth muscle cells senescence and inflammation in aged mouse aortas [76]. Similarly, miRNA-217 was identified as an endogenous SIRT1 inhibitor, which promotes endothelial senescence. MiRNA-217 was expressed in human atherosclerotic lesions and was negatively correlated with SIRT1 expression and with FoxO1 acetylation status [77]. Zhao et al. showed that miRNA-128-3p aggravated the doxorubicin-induced liver injury by promoting oxidative stress via targeting SIRT1 [78]. The study of Zhu et al. demonstrates a pro-apoptotic role of miRNA-195 in cardiomyocytes and identifies SIRT1 as a direct target of miRNA-195. The effect of miRNA-195 on apoptosis is mediated through the downregulation of SIRT1, Bcl-2 (B-cell lymphoma 2), and ROS production [79]. D'Adamo et al. identified miRNA-9 as a post-transcriptional regulator of SIRT1. MiRNA-9 and SIRT1 levels showed opposite changes in chondrocytes, following H₂O₂ treatment [80].

NF-κB Pathway

Excessive levels of ROS would also activate NF-κB pathway that is a redox-sensitive pathway [81]. NF-κB is present in all kinds of cells controlling the transcription of a wide variety of genes, including pro-apoptotic and pro-survival genes, pro-inflammatory cytokines, antioxidant and pro-oxidant enzymes, and many others [82]. The mammalian NF-κB family is composed of five members: p65 (RelA), RelB, c-Rel, NF-κB1 (p50 and its precursor p105), and NF-κB2 (p52 and its precursor p100), which can form homodimers and heterodimers among themselves. The NF-κB proteins are normally sequestered in the cytoplasm by a family of inhibitory proteins, including IκB family members. The most common activation reactions of NF-κB are represented by phosphorylation and activation of the IκB kinase complex [83].

One of the most important ways in which NF-κB activity influences ROS levels is via increased expression of antioxidant proteins such as SOD, glutathione peroxidase, or heme oxygenase. Since NF-κB is important in inflammation, some

enzymes that promote the production of ROS (e.g., NOX2, inducible nitric oxide synthase (iNOS), cyclooxygenase (COX) 2, or cytochrome P450 enzymes) are also regulated as its targets, especially in cells of the immune system [82]. Abnormal NF- κ B activity is frequently associated with an abnormal level of miRNAs, which is found to play critical roles in disease progression [84].

Downregulation of miRNA-155 ameliorates high-glucose-induced endothelial injury by inhibiting NF- κ B activation and promoting HO-1 and NO production [71]. Gu revealed that miRNA-124 prevents H₂O₂-induced oxidative stress and apoptosis in human lens epithelial cells by suppressing the activation of the NF- κ B pathway [85]. Results of Wei et al. indicated that NF- κ B positively regulated miRNA-21 expression under oxidative stress, and programmed cell death protein 4 (PDCD4) was a direct target for miRNA-21 [86]. In another study, Xie et al. studied the role of miRNA-146a in the brain of chronic type 2 diabetes mellitus (cT2DM) rats. Increased inflammation and oxidative stress were associated with brain impairment in cT2DM rats, which were negatively correlated with miR-146a expression. The expressions of NF- κ B p65 and its specific modulators were elevated in the brain of cT2DM rats, which might be inhibited by miR-146a [87].

Several other genes and related pathways have been described in the literature to be involved in the regulation of oxidative stress by miRNAs. Examples of these are MAPK (mitogen-activated protein kinase) signaling pathway, TGF- β (transforming growth factor beta) signaling pathway, cell adhesion molecules (CAMs), calcium signaling pathway, VEGF (vascular endothelial growth factor) signaling pathway, etc. [61].

MiRNA in Oxidative-Stress-Induced Heart Diseases

Oxidative stress plays a crucial role in many cardiovascular diseases, like hypoxia, ischemia/reperfusion injury, or heart failure [88]. As mentioned before, intracellular ROS are formed in normal conditions as the result of normal mitochondrial

respiration, but ROS are also produced during reperfusion in hypoxic tissue and in association with infection and inflammation, leading to pathological conditions of the heart [88,89]. One of the effects of ROS accumulation in cardiomyocytes is a different expression of noncoding RNAs (ncRNAs), subsequently contributing to cell apoptosis and heart pathology. Among these ncRNAs, miRNAs are the most intensively studied, as they have a huge impact on heart condition by inhibiting protein translation or target mRNA degradation [16,90,91].

Cardiac Hypertrophy

Oxidative stress generates many complex cellular changes in the heart which force it to adaptation in the form of cardiac hypertrophy of ventricles. These adaptations may provide initial salutary compensation to the arisen stress, sustained hypertrophic stimulation becomes maladaptive, worsening morbidity, and mortality risks because of congestive heart failure and sudden cardiac death [92]. During hypertrophy in different experiments with animal models or in clinical trials, changes in miRNA expression were observed—mainly miRNA-1 and -133. Zhao et al. demonstrated that their lower levels indicate a significant cardiac injury [93]. MiRNA-1 is connected with cardiomyocyte growth and hypertrophy most probably through the calcineurin/nuclear factor of activated T cell (NFAT) signaling pathway inhibition [94,95]. As a previous miRNA-1 case, also patients and animals with cardiac hypertrophy have lower levels of miRNA-133, probably by regulating antihypertrophic genes like guanosine diphosphate–guanosine triphosphate (GDP–GTP) exchange protein, or signal transduction kinase cell division control protein 42 (Cdc42) [96,97].

MiRNA-208 belongs to the cardiac-specific miRNAs, and its regulation is important in the processes of cardiac remodeling. Several studies demonstrated that cardiac hypertrophy is caused by switching of adult alpha myosin heavy chain (α -MHC, known as Myh6) to fetal beta myosin heavy chain (β -MHC, known as Myh7) gene expression. In experimental studies, deletion of miRNA-208a, which is encoded in the intron of the Myh6 gene,

leads to the decreased expression of the Myh7 gene in response to stress and to hypertrophy [88,98]. These results were confirmed in the study of Rawal et al., where inhibition of miRNA-208a hampers the activation of β -MHC and hypertrophic response [99].

Important miRNAs involved in the cardiac hypertrophy are also miRNA-22 (influence phosphatidylinositol-3-kinase (PI3K)-protein kinase B (AKT)) [100], miRNA-212/132 family (active through antihypertrophic FoxO3 transcription factor), or miRNA-199 (miRNA-199a targets the pro-autophagic and antihypertrophic factor glycogen synthase kinase 3 β ; miRNA-199b acts through targeting tyrosine phosphorylation regulated kinase 1A (Dyrk1a) gene, involved in the phosphorylation of NFAT factors) [101-103]. Other studies dealing with miRNAs associated with cardiac hypertrophy observed changed expression of miRNA-21, -18b, -195, -199, -29, -22, or -23 levels [104-107].

Ischemia/Reperfusion Injury

Cardiac ischemia/reperfusion (I/R) injury involves the damage caused by reduced coronary blood flow, causing depletion of ATP, reduced partial pressure of oxygen, and production of toxins. Reperfusion leads to further damage through generation of ROS and a proton gradient across both the sarcolemma and the inner mitochondrial membrane [108]. Many miRNAs are involved in these processes, either as a result of damage caused by ROS generation or directly responsive to ROS.

One of the most promising miRNAs for potential use as a diagnostic or therapeutic target is miRNA-24-3p. In a very recent study of Xiao et al., decrease expression of miRNA-24-3p during induced ischemia/reperfusion injury in mouse hearts and decreasing levels of apoptosis of cardiomyocytes caused by ROS during ischemia/reperfusion injury after application of miRNA mimics were confirmed [109]. In addition to these observations, the authors identified the Keap1-Nrf2 pathway as one of the possible targets of miRNA-24-3p [109].

Lusha et al. showed that miRNA-144, which is primarily connected with the regulation of apoptosis in human cancer diseases, is another miRNA with changed expression levels in the I/R model through regulation of FoxO1. The authors observed in their study reduced infarct size and apoptosis in cardiomyocytes during the upregulation of miRNA-144 and increased sensitivity to I/R in the situation with depleted miRNA-144 [110]. FoxO1 protein is an important transcription factor which mediates apoptosis by activating iNOS expression in cardiomyocytes [111], and it can be regulated by sirtuin 1 in the cardiovascular system [112].

In another study, Fang and Yeh explored the role of miRNAs in cardiomyocyte apoptosis induced by ischemia. They found an increased level of miRNA-302 expression induced by hypoxia/reoxygenation injury. This aggravated cardiomyocyte apoptosis probably by inhibiting antiapoptotic protein myeloid cell leukemia 1 (Mcl-1) expression, thereby activating pro-apoptotic molecules. Based on this data, the authors suggested that elevated miRNA-302 levels can be detrimental to cells, but decreased levels are beneficial and can lead to effective therapeutic intervention [113]. Several publications reveal profound effects of myocardial ischemia on miRNA transcript in the vessel wall and vascular smooth muscle cells, in particular [114-116].

MiRNA-23a promotes cardiomyocyte apoptosis and myocardial infarction induced by I/R through directly suppressing the expression of manganese SOD, an important antioxidant for scavenging of superoxide [117]. This is one of the endogenous enzymes that protects cells from oxidative stress. Wang et al. observed that miRNA-1 worsens cardiac oxidative stress by post-transcriptional modification of the antioxidant network in the I/R injury C57BL/6 mice model. They found that miRNA-1 reduced the protein levels of antioxidant enzymes glutamate cysteine ligase (Gclc), SOD1, and glucose-6-phosphate dehydrogenase (G6PD) under oxidative stress conditions [118]. Other miRNAs, such as miRNA-130a [119] and miRNA-98 [120], are investigated to be associated with ROS-related cardiomyocyte apoptosis. MiRNA-208a promoted apoptosis and

oxidative stress in the I/R injury rat model by regulation of protein tyrosine phosphatase receptor type G and protein tyrosine phosphatase, non-receptor type 4 [121].

Another situation of ischemia/reperfusion injury is heart transplantations. During the transplantation process in the final step, where heart graft is connected to the circulatory system of the recipient and when reperfusion of the graft is started, there is an excessive production of free radicals, what could lead to graft failures and lower long-term survival rate of the patients [122]. In several works, more attention is focused on miRNA's changed expression after transplantation, making miRNAs as an ideal candidate for biomarkers of transplant rejection [122-124]. Zhou et al. found upregulation of miRNA-711, -2137, -705, -5130, -346, -714, and -744 and downregulation of miRNA-210, -490, -491, -425, -423-3p, and -532-3p in experiments on C56BL/6 mouse animal models after heart transplantation in I/R injured hearts [122].

MiRNAs were measured in the samples from endomyocardial biopsies (EMB) and blood serum in 30 patients with rejecting heart allograft and 30 patients without allograft rejection. MiRNA analyzes revealed changed expression of miRNA-10a, -31, -92a, and -155 in EMB, as well as in blood serum [125]. These miRNAs are strongly associated with inflammatory processes, as they influenced NF- κ B, TNF- α , interleukins -6, -8, and -1, monocyte chemoattractant protein-1, eNOS, or vascular cell adhesion protein [126-128]. Wei et al. also suggested that miRNA-183, -182, and -96 have an important function in the regulation of graft rejection probably through regulation of FoxO1 expression and could be new potential noninvasive biomarkers of allograft rejection in heart transplantation [129].

All these results suggested the potential role of miRNAs in the regulation and adaptation of transplanted allografts and their potential use as biomarkers in grafts rejection.

Coronary Artery Diseases (CAD)

CAD is an atherosclerotic disease which is inflammatory in nature. Atherosclerosis starts due to the accumulation of

lipoproteins in the intima of the coronary vessels. Oxidized or modified low-density lipoprotein then attract leukocytes into the intima of the coronary vessels, which can be scavenged by macrophages, leading to the formation of foamy cells. The atherosclerotic plaque starts developing. Cell death or apoptosis occurs commonly in the atherosclerotic lesions. The modified lipoproteins propagate inflammatory responses. As a result, obstruction of blood flow occurs, and this leads to a mismatch between myocardial oxygen demand and supply [130].

Endothelial cell apoptosis under oxidative stress plays a critical role in the initiation and progression of atherosclerosis [131-141]. Li et al. observed that overexpression of miRNA-210 caused inhibition of apoptosis and reduction of ROS level in human umbilical vein endothelial cells (HUVECs) treated with H₂O₂ and also downregulation of caspase levels. This study leads to the conclusion that miRNA-210 could have a role in the protection against oxidative-stress-induced apoptosis in HUVECs [131].

MiRNA-24 is highly expressed in the vessel wall and changes of its expression are connected with dysfunction and injury of vascular endothelial cells [132]. It is believed that miRNA-24 participates in many pathophysiological processes including I/R injury or vascular oxidative stress [134-137]. In an experimental study by Zhang et al., miRNA-24 was upregulated, and it has a supportive effect on vascular endothelium repair by attenuating oxidative-stress-induced damage of endothelial cells. In this case, miRNA-24 regulated the Nrf2/HO-1 signaling pathway indirectly by regulating of Keap1 after influencing of O-linked β -N-acetylglucosamine transferase gene (Ogt). On the other hand, authors confirmed that miRNA-24 affects the expression of SOD, malondialdehyde, and glutathione peroxidase [137].

MiRNA-92a overexpression impairs endothelial function and suppresses HO-1 expression in endothelial cells. Inhibition of miRNA-92a attenuates oxidative stress and improves endothelial function through enhancing HO-1 expression and activity in diabetic mouse aortas [138]. Yamac et al. observed markedly lowered expression of miRNA-199a in patients with coronary

artery disease [139]. In parallel, they also showed the induction of cardioprotective protein SIRT1, a potential target of miRNA-199. Significantly increased expression of miRNA-146a was observed in patients with acute coronary syndrome. This miRNA is connected with the inflammatory pathway by regulating NF- κ B [140]. According to O'Sullivan et al., miRNA-93-5p belongs to one of the strongest predictors of coronary artery disease when its expression was significantly upregulated in the patients with CAD, most likely through modulation of ATP-binding cassette A1 (ABCA1) gene [141]. ABCA1 plays an important role in cholesterol homeostasis and atherogenesis, and it could be reduced by oxidative stress [133].

Heart Failure

A complex syndrome resulting from structural or functional cardiac disorders, leading to disability of ventricle to fill or eject blood, is called heart failure [142-144], and it is considered to be one of the leading cause of morbidity and mortality worldwide [142,145-148]. Development of heart failure depends on many circumstances in the organism, but one of the key pathophysiological pathways for it is oxidative stress [149-151]. According to a lot of animal and human studies, multiple miRNAs are changed in models of heart failure, including miRNA-199b, -195, -100, -133, -24, and -208 [94,152-156]. Upregulated levels of miRNA-199b were measured during heart failure and appeared to target the calcineurin/NFAT pathway. It was proved that this calcineurin/NFAT pathway is activated after oxidative stress stimuli [157,158]. In the in vivo experiments, inhibition of miRNA-199b caused normalization of the expression of Dyrk1a, a reduction of nuclear NFAT activity, and inhibition of hypertrophy and fibrosis in mouse models of heart failure [101,159]. Changed expressions of miRNA-1, -214, -29b, -342, -7, -107, -126, -125, -122, -423-5p, -320a, -650, -1228, -662, -583, -3175, -21, -22, and -92b have been shown in other studies of heart failure [106,152,156,160-163].

A brief review of selected miRNAs included in the cardiovascular diseases caused by oxidative stress is provided in Table 1.

Table 1: List of selected miRNAs with function in oxidative-stress-induced cardiovascular diseases.

Disease	miRNA	Expression	Target	References
Cardiac hypertrophy	miRNA-1	Downregulated	Mef2a; Gata4	[93,94,95]
	miRNA-133	Downregulated	GDP-GTP exchange protein; Cdc42	[93,96,97]
	miRNA-208a	Downregulated	<i>Myh7</i>	[18,98,99]
Ischemia/reperfusion injury	miRNA-24-3p	Downregulated	Keap1/Nrf2	[109]
	miRNA-144	Downregulated	FoxO1	[110,111]
	miRNA-302	Upregulated	Mcl-1	[113]
	miRNA-23a	Upregulated	MnSOD	[117]
Heart transplantation	miRNA-10a	Downregulated	NF-κB	[126]
	miRNA-31	Upregulated	TNF-α	[127]
	miRNA-92a	Upregulated	Integrin α5, S1P1, MKK4, eNOS	[128]
	miRNA-155	Upregulated	T-cell receptor, IFN receptor	[164]
Coronary artery diseases	miRNA-24	Downregulated	<i>Ogt</i> , Keap1/Nrf2	[132,134,135,136,137]
	miRNA-92a	Upregulated	HO-1	[138]
	miRNA-199a	Downregulated	SIRT1	[139]
Heart failure	miRNA-199b	Upregulated	calcineurin/NFAT	[101,157,158,159]
	miRNA-21	Upregulated	natriuretic peptide B	[156,163]

Mef2a—Myocyte-specific enhancer factor 2A, Gata4—Transcription factor GATA-4, GDP-GTP exchange protein—guanosine diphosphate-guanosine triphosphate exchange protein, Cdc42—Cell division control protein 42, *Myh7*—beta myosin heavy chain, Keap1/Nrf2—Kelch-like ECH-associated protein 1/Nuclear factor erythroid 2-related factor 2, FoxO1—Forkhead box protein O1, Mcl-1—Myeloid cell leukemia 1, MnSOD—manganese-dependent superoxide dismutase, NF-κB—nuclear factor kappa B, TNF-α—tumor necrosis factor alpha, S1P1—Sphingosine-1-phosphate receptor 1, MKK4—Mitogen-activated protein kinase kinase 4, eNOS—Endothelial nitric oxide synthase, INF receptor—interferon receptor, *Ogt*—O-linked β-N-acetylglucosamine transferase, HO-1—Heme oxygenase 1, SIRT1—sirtuin 1, NFAT—Nuclear factor of activated T cells.

Future Perspectives of Using MiRNA in Disease Diagnosis and Treatment

Since 2001, miRNAs have been recognized as biomarkers and possible therapeutic targets for the diagnosis and treatment of diseases [165]. One of the biggest advantages for using miRNAs as biomarkers is their stability under many different conditions. MiRNAs can be stored at room temperature, frozen, or thawed [166]. Bioavailability of miRNA is another great advantage. MiRNAs can be isolated from various biological materials, like from peripheral blood, fresh and frozen tissues, or formalin-fixed, paraffin-wax-embedded samples, but also from saliva, epithelium of the skin, or hair [167,168]. Difficulties in the use of therapeutically altering miRNAs lie in their non-specificity—single miRNA can target many genes and influence more than one gene expression, so they could affect also other pathways in the organisms [169]. MiRNAs impose a relatively modest effect on their target, reflecting that individual mRNAs are targeted by multiple miRNAs, while the cellular proteome might be able to compensate the absence of a single miRNA [170].

Manipulation of RNA using miRNA mimics and antagomirs holds significant therapeutic potential for treating a variety of diseases. With recent technological advances, identification and validation of potential therapeutic miRNA targets are readily available [165]. Treatment of diseases by modulation of selected miRNAs in the organisms is based on two approaches. First, miRNA mimics is an approach for gene silencing due to generating synthesized artificial double-stranded miRNA-like RNA fragments. These molecules are able to bind to target mRNA and suppressed genes [171]. The second approach uses antagomirs, chemically designed oligonucleotides. These oligonucleotides specifically inhibit target miRNA by binding to them, which leads to reduction of RISC activation and to upregulation of genes [172,173]. MiRNAs could be modulated also by miRNA sponges (target mimicry), masking, and erasers. MiRNA sponges contain a binding site for the miRNA family, leading to the blocking of the activity of miRNAs [174,175]. Masking is based on the occupation of the binding site on target mRNA by oligonucleotides [176]. Erasers are oligonucleotides

complementary to specific miRNA, leading to inhibition of its function [177]. However, delivery of anti-miRNAs and miRNAs in vivo may prove to be challenging [165].

Conclusions

Oxidative stress is one of the important contributing factors in cardiovascular disease genesis and development. Excessive ROS production has a significant impact on the pathogenesis of cardiovascular diseases related to atherosclerosis, cardiomyopathy, ischemia/reperfusion, and heart failure. Published literature highlights the increasing importance of studying the role of redox-sensitive miRNAs to identify more effective biomarkers and develop better therapeutic targets for oxidative-stress-related diseases. It is necessary to define the roles of individual miRNAs and their important targets, to determine their potential for possible diagnosis/treatment of cardiovascular disorders. Although a number of targets of oxidative-stress-responsive miRNAs have been identified, e.g., Nrf2, SIRT1, and NF- κ B, future studies are still needed to determine further potential targets and their links to cardiovascular disease. MiRNA may be a promising novel tool and means in the clinical diagnosis, prognostic evaluation, and even therapeutic intervention of oxidative-stress-related CVD. The knowledge of the crosstalk between miRNAs, ROS, and cardiovascular diseases can contribute to new therapeutic approaches based on the suppression of ROS effects, with the potential to ameliorate or prevent the progression of cardiovascular diseases. However, several studies are still required to validate the present findings before the application of miRNA in clinical practice.

References

1. Roth GA, Abate D, Abate KH, Abay SM, Abbafati C, et al. Global, regional, and national age-sex-specific mortality for 282 causes of death in 195 countries and territories, 1980–2017: A systematic analysis for the Global Burden of Disease Study 2017. *Lancet*. 2018; 392: 1736–1788.

2. WHO. About Cardiovascular Diseases. Available online at: https://www.who.int/cardiovascular_diseases/about_cvd/en/
3. Li M, Duan L, Li Y, Liu B. Long noncoding RNA/circular noncoding RNA–miRNA–mRNA axes in cardiovascular diseases. *Life Sci.* 2019; 233: 116440.
4. Hansen TB, Jensen TI, Clausen BH, Bramsen JB, Finsen B, et al. Natural RNA circles function as efficient microRNA sponges. *Nature.* 2013; 495: 384–388.
5. Honda T, Hirakawa Y, Nangaku M. The role of oxidative stress and hypoxia in renal disease. *Kidney Res. Clin. Pract.* 2019; 38: 414.
6. Zhang J, Wang X, Vikash V, Ye Q, Wu D, et al. ROS and ROS-Mediated Cellular Signaling. *Oxid. Med. Cell. Longev.* 2016; 2016: 4350965.
7. Sugamura K, Keaney JF. Reactive oxygen species in cardiovascular disease. *Free Radic. Biol. Med.* 2011; 51: 978–992.
8. Sawyer DB, Siwik DA, Xiao L, Pimentel DR, Singh K, Colucci WS. Role of oxidative stress in myocardial hypertrophy and failure. *J. Mol. Cell. Cardiol.* 2002; 34: 379–388.
9. Wongsurawat T, Woo CC, Giannakakis A, Lin XY, Cheow ESH, et al. Transcriptome alterations of vascular smooth muscle cells in aortic wall of myocardial infarction patients. *Data Br.* 2018; 17: 1112–1135.
10. Wongsurawat T, Woo CC, Giannakakis A, Lin XY, Cheow ESH, et al. Distinctive molecular signature and activated signaling pathways in aortic smooth muscle cells of patients with myocardial infarction. *Atherosclerosis.* 2018; 271: 237–244.
11. Derda AA, Woo CC, Wongsurawat T, Richards M, Lee CN, et al. Gene expression profile analysis of aortic vascular smooth muscle cells reveals upregulation of cadherin genes in myocardial infarction patients. *Physiol. Genom.* 2018; 50: 648–657.
12. Date RA. Bradyrhizobium effectiveness responses in *Stylosanthes hamata* and *S. seabrana*. *Trop. Grassl.* 2010; 44: 141–157.
13. Ambros V. microRNAs: Tiny regulators with great potential. *Cell.* 2001; 107: 823–826.

14. Treiber T, Treiber N, Meister G. Regulation of microRNA biogenesis and its crosstalk with other cellular pathways. *Nat. Rev. Mol. Cell Biol.* 2019; 20: 5–20.
15. Pasquinelli AE. MicroRNAs and their targets: Recognition, regulation and an emerging reciprocal relationship. *Nat. Rev. Genet.* 2012; 13: 271–282.
16. Filipowicz W, Bhattacharyya SN, Sonenberg N. Mechanisms of post-transcriptional regulation by microRNAs: Are the answers in sight? *Nat. Rev. Genet.* 2008; 9: 102–114.
17. Duygu B, de Windt LJ, da Costa Martins PA. Targeting microRNAs in heart failure. *Trends Cardiovasc. Med.* 2016; 26: 99–110.
18. Colpaert RMW, Calore M. MicroRNAs in Cardiac Diseases. *Cells.* 2019; 8: 737.
19. Valášková Z, Mladosičičová B, Hulín I, Maruščáková L. MicroRNA—information about myocardial damage or biomarker of heart failure? *Cardiol. Lett.* 2017; 26: 299–302.
20. Kreutzer FP, Fiedler J, Thum T. Non-coding RNAs: Key players in cardiac disease. *J. Physiol.* 2019.
21. Chistiakov DA, Orekhov AN, Bobryshev YV. Cardiac-specific miRNA in cardiogenesis, heart function, and cardiac pathology (with focus on myocardial infarction). *J. Mol. Cell. Cardiol.* 2016; 94: 107–121.
22. Deng J, Zhong Q. Advanced research on the microRNA mechanism in heart failure. *Int. J. Cardiol.* 2016; 220: 61–64.
23. Gong YY, Luo JY, Wang L, Huang Y. MicroRNAs Regulating Reactive Oxygen Species in Cardiovascular Diseases. *Antioxid. Redox Signal.* 2018; 29: 1092–1107.
24. Münzel T, Camici GG, Maack C, Bonetti NR, Fuster V, et al. Impact of Oxidative Stress on the Heart and Vasculature Part 2 of a 3-Part Series HHS Public Access Pathophysiological Role of Oxidative Stress in Heart Failure. *J. Am. Coll. Cardiol.* 2017; 70: 212–229.
25. Akhtar MJ, Ahamed M, Alhadlaq HA, Alshamsan A. Mechanism of ROS scavenging and antioxidant signalling by redox metallic and fullerene nanomaterials: Potential implications in ROS associated degenerative disorders. *Biochim. Biophys. Acta.* 2017; 1861: 802–813.

26. Palade F, Alexa ID, Azoicai D, Panaghiu L, Ungureanu G. Oxidative stress in atherosclerosis. *Rev. Med.* 2003; 107: 502–511.
27. Halliwell B. Antioxidants in Human Health and Disease. *Annu. Rev. Nutr.* 1996; 16: 33–50.
28. Kurian GA, Rajagopal R, Vedantham S, Rajesh M. The Role of Oxidative Stress in Myocardial Ischemia and Reperfusion Injury and Remodeling: Revisited. *Oxid. Med. Cell. Longev.* 2016; 2016: 1656450.
29. Syu JP, Chi JT, Kung HN. Nrf2 Contributes to the Poor Prognosis and Chemoresistance. *A Master Regul. Oxidative Stress.* 2016.
30. Finkel T. Signal transduction by reactive oxygen species. *J. Cell Biol.* 2011; 194: 7–15.
31. Mccord J, Fridovich I. Superoxide dismutase. An enzymic function for erythrocyte hemocuprein (hemocuprein). *J. Biol. Chem.* 1969; 244: 6049–6055.
32. Wang Y, Branicky R, Noë A, Hekimi S. Superoxide dismutases: Dual roles in controlling ROS damage and regulating ROS signaling. *J. Cell Biol.* 2018; 217: 1915–1928.
33. Li H, Horke S, Förstermann U. Oxidative stress in vascular disease and its pharmacological prevention. *Trends Pharmacol. Sci.* 2013; 34: 313–319.
34. Cheung O, Sanyal AJ. MicroRNAs. *Signal. Pathw. Liver Dis.* 2010; 107: 493–499.
35. Moris D, Spartalis M, Spartalis E, Karachaliou GS, Karaolani GI, et al. The role of reactive oxygen species in the pathophysiology of cardiovascular diseases and the clinical significance of myocardial redox. *Ann. Transl. Med.* 2017; 5: 326.
36. Gori T, Münzel T. Oxidative stress and endothelial dysfunction: Therapeutic implications. *Ann. Med.* 2011; 43: 259–272.
37. Maack C, Kartes T, Kilter H, Schäfers HJ, Nickenig G, et al. Oxygen free radical, release in human failing myocardium is associated with increased activity of Rac1-GTPase and represents a target for statin treatment. *Circulation.* 2003; 108: 1567–1574.

38. Mollnau H, Oelze M, August M, Wendt M, Daiber A, et al. Mechanisms of increased vascular superoxide production in an experimental model of idiopathic dilated cardiomyopathy. *Arterioscler. Thromb. Vasc. Biol.* 2005; 25: 2554–2559.
39. Bendall JK, Cave AC, Heymes C, Gall N, Shah AM. Pivotal role of a gp91phox-containing NADPH oxidase in angiotensin II-induced cardiac hypertrophy in mice. *Circulation* 2002, 105, 293–296.
40. Nakagami, H., Takemoto, M., Liao, J.K. NADPH oxidase-derived superoxide anion mediates angiotensin II-induced cardiac hypertrophy. *J. Mol. Cell. Cardiol.* 2003; 35: 851–859.
41. Wenzel P, Knorr M, Kossmann S, Stratmann J, Hausding M, et al. Lysozyme M-positive monocytes mediate angiotensin ii-induced arterial hypertension and vascular dysfunction. *Circulation.* 2011; 124: 1370–1381.
42. Ago T, Kuroda J, Pain J, Fu C, Li H, et al. Upregulation of Nox4 by hypertrophic stimuli promotes apoptosis and mitochondrial dysfunction in cardiac myocytes. *Circ. Res.* 2010; 106: 1253–1264.
43. Kuroda J, Ago T, Matsushima S, Zhai P, Schneider MD, et al. NADPH oxidase 4 (Nox4) is a major source of oxidative stress in the failing heart. *Proc. Natl. Acad. Sci. USA.* 2010; 107: 15565–15570.
44. Li J, Romestaing C, Han X, Li Y, Hao X, et al. Cardiolipin remodeling by ALCAT1 links oxidative stress and mitochondrial dysfunction to obesity. *Cell Metab.* 2010; 12: 154–165.
45. Gadicherla AK, Stowe DF, Antholine WE, Yang M, Camara AKS. Damage to mitochondrial complex i during cardiac ischemia reperfusion injury is reduced indirectly by anti-anginal drug ranolazine. *Biochim. Biophys. Acta.* 2012; 1817: 419–429.
46. Paradies G, Petrosillo G, Pistolese M, Ruggiero FM. Reactive oxygen species affect mitochondrial electron transport complex I activity through oxidative cardiolipin damage. *Gene.* 2002; 286: 135–141.
47. Baldus S, Müllerleile K, Chumley P, Steven D, Rudolph V, et al. Inhibition of xanthine oxidase improves myocardial

- contractility in patients with ischemic cardiomyopathy. *Free Radic. Biol. Med.* 2006; 41: 1282–1288.
48. Touyz RM, Chen X, Tabet F, Yao G, He G, et al. Expression of a functionally active gp91phox-containing neutrophil-type NAD(P)H oxidase in smooth muscle cells from human resistance arteries: Regulation by angiotensin II. *Circ. Res.* 2002; 90: 1205–1213.
 49. Bitar MS, Wahid S, Mustafa S, Al-Saleh E, Dhaunsi GS, et al. Nitric oxide dynamics and endothelial dysfunction in type II model of genetic diabetes. *Eur. J. Pharmacol.* 2005; 511: 53–64.
 50. Pedro-Botet J, Covas MI, Martín S, Rubiés-Prat J. Decreased endogenous antioxidant enzymatic status in essential hypertension. *J. Hum. Hypertens.* 2000; 14: 343–345.
 51. Münzel T, Gori T, Bruno RM, Taddei S. Is oxidative stress a therapeutic target in cardiovascular disease? *Eur. Heart J.* 2010; 31: 2741–2749.
 52. Shi S, Xue F. Current antioxidant treatments in organ transplantation. *Oxid. Med. Cell. Longev.* 2016; 2016: 8678510.
 53. Vela C, Thomsen M, Delbosc S, Calise D, Cristol JP, et al. Lipid and Oxidative Stress Disorders in a Rat Model of Chronic Rejection. *Transplant. Proc.* 2007; 39: 2617–2619.
 54. Larsen M, Webb G, Kennington S, Kelleher N, Sheppard J, et al. Mannitol in cardioplegia as an oxygen free radical scavenger measured by malondialdehyde. *Perfusion.* 2002; 17: 51–55.
 55. Rosenkranz ER. Substrate enhancement of cardioplegic solution: Experimental studies and clinical evaluation. *Ann. Thorac. Surg.* 1995; 60: 797–800.
 56. Fudulu D, Angelini G. Oxidative Stress after Surgery on the Immature Heart. *Oxid. Med. Cell. Longev.* 2016; 2016: 1971452.
 57. Ryan JB, Hicks M, Cropper JR, Nicholson A, Kesteven SH, et al. Lazaroid (U74389G)-supplemented cardioplegia: Results of a double-blind, randomized, controlled trial in a porcine model of orthotopic heart transplantation. *J. Hear. Lung Transplant.* 2003; 22: 347–356.
 58. Watson AJ, Gao L, Sun L, Tsun J, Jabbour A, et al. Enhanced preservation of the rat heart after prolonged

- hypothermic ischemia with erythropoietin-supplemented Celsior solution. *J. Hear. Lung Transplant.* 2013; 32: 633–640.
59. Villanueva JE, Gao L, Chew HC, Hicks M, Doyle A, et al. Functional recovery after dantrolene supplementation of cold stored hearts using an ex vivo isolated working rat heart model. *PLoS ONE.* 2018; 13: e0205850.
 60. Schieber M, Chandel NS. ROS function in redox signaling and oxidative stress. *Curr. Biol.* 2014; 24: R453–R462.
 61. Engedal N, Žerovnik E, Rudov A, Galli F, Olivieri F, et al. From oxidative stress damage to pathways, networks, and autophagy via microRNAs. *Oxid. Med. Cell. Longev.* 2018; 2018: 4968321.
 62. Wan Y, Cui R, Gu J, Zhang X, Xiang X, et al. Identification of Four Oxidative Stress-Responsive MicroRNAs, miR-34a-5p, miR-1915-3p, miR-638, and miR-150-3p, in Hepatocellular Carcinoma. *Oxid. Med. Cell. Longev.* 2017; 2017: 5189138.
 63. Banerjee J, Khanna S., Bhattacharya, A. MicroRNA Regulation of Oxidative Stress. *Oxid. Med. Cell. Longev.* 2017; 2017: 2872156.
 64. Chen JY, Zhu GY, Su XH, Wang R, Liu J, et al. 7-deacetylgedunin suppresses inflammatory responses through activation of Keap1/Nrf2/HO-1 signaling. *Oncotarget.* 2017; 8: 55051–55063.
 65. Chen B, Lu Y, Chen Y, Cheng J. The role of Nrf2 in oxidative stress-induced endothelial injuries. *J. Endocrinol.* 2015; 225: R83–R99.
 66. Zhu J, Wang S, Qi W, Xu X, Liang Y. Overexpression of miR-153 promotes oxidative stress in MPP⁺-induced PD model by negatively regulating the Nrf2/HO-1 signaling pathway. *Int. J. Clin. Exp. Pathol.* 2018; 11: 4179–4187.
 67. Sangokoya C, Telen MJ, Chi JT. microRNA miR-144 modulates oxidative stress tolerance and associates with anemia severity in sickle cell disease. *Blood.* 2010; 116: 4338–4348.
 68. Yin Y, Zhao X, Yang Z, Min X. Downregulation of miR-93 elevates Nrf2 expression and alleviates reactive oxygen species and cell apoptosis in diabetic retinopathy. *Int. J. Clin. Exp. Med.* 2019; 12: 10235–10243.

69. Cheng LB, Li K, Yi N, Li XM, Wang F, et al. miRNA-141 attenuates UV-induced oxidative stress via activating Keap1-Nrf2 signaling in human retinal pigment epithelium cells and retinal ganglion cells. *Oncotarget*. 2017; 8: 13186–13194.
70. Kabaria S, Choi DC, Chaudhuri AD, Jain MR, Li H, et al. MicroRNA-7 activates Nrf2 pathway by targeting Keap1 expression. *Free Radic. Biol. Med.* 2015; 89: 548–556.
71. Zhang S, Wu W, Jiao G, Li C, Liu H. MiR-455-3p activates Nrf2/ARE signaling via HDAC2 and protects osteoblasts from oxidative stress. *Int. J. Biol. Macromol.* 2018; 107: 2094–2101.
72. Yang JJ, Tao H, Hu W, Liu LP, Shi KH, et al. MicroRNA-200a controls Nrf2 activation by target Keap1 in hepatic stellate cell proliferation and fibrosis. *Cell. Signal.* 2014; 26: 2381–2389.
73. Kwon J, Lee S, Kim YN, Lee IH. Deacetylation of CHK2 by SIRT1 protects cells from oxidative stress-dependent DNA damage response. *Exp. Mol. Med.* 2019; 51: 36.
74. Salminen A, Kaarniranta K, Kauppinen A. Crosstalk between oxidative stress and SIRT1: Impact on the aging process. *Int. J. Mol. Sci.* 2013; 14: 3834–3859.
75. Sengupta A, Molkentin JD, Paik JH, DePinho RA, Yutzey KE. FoxO transcription factors promote cardiomyocyte survival upon induction of oxidative stress. *J. Biol. Chem.* 2011; 286: 7468–7478.
76. Badi I, Burba I, Ruggeri C, Zeni F, Bertolotti M, et al. MicroRNA-34a Induces Vascular Smooth Muscle Cells Senescence by SIRT1 Downregulation and Promotes the Expression of Age-Associated Pro-inflammatory Secretory Factors. *J. Gerontol.* 2015; 70: 1304–1311.
77. Menghini R, Casagrande V, Cardellini M, Martelli E, Terrinoni A, et al. MicroRNA 217 modulates endothelial cell senescence via silent information regulator 1. *Circulation.* 2009; 120: 1524–1532.
78. Zhao X, Jin Y, Li L, Xu L, Tang Z, et al. MicroRNA-128-3p aggravates doxorubicin-induced liver injury by promoting oxidative stress via targeting Sirtuin-1. *Pharmacol. Res.* 2019; 146: 104276.
79. Zhu H, Yang Y, Wang Y, Li J, Schiller PW, et al. MicroRNA-195 promotes palmitate-induced apoptosis in

- cardiomyocytes by down-regulating Sirt1. *Cardiovasc. Res.* 2011; 92: 75–84.
80. D’Adamo S, Cetrullo S, Guidotti S, Borzì RM, Flamigni F. Hydroxytyrosol modulates the levels of microRNA-9 and its target sirtuin-1 thereby counteracting oxidative stress-induced chondrocyte death. *Osteoarthr. Cartil.* 2017; 25: 600–610.
 81. Gaspar-Pereira S, Fullard N, Townsend PA, Banks PS, Ellis EL, et al. The NF- κ B subunit c-Rel stimulates cardiac hypertrophy and fibrosis. *Am. J. Pathol.* 2012; 180: 929–939.
 82. Morgan MJ, Liu ZG. Crosstalk of reactive oxygen species and NF- κ B signaling. *Cell Res.* 2011; 21: 103–115.
 83. Ma X, Becker Buscaglia LE, Barker JR, Li Y. MicroRNAs in NF- κ B signaling. *J. Mol. Cell Biol.* 2011; 3: 159–166.
 84. Yuan Y, Tong L, Wu S. MicroRNA and NF-kappa B. *Adv. Exp. Med. Biol.* 2015; 887: 157–170.
 85. Gu XL. MicroRNA-124 Prevents H₂O₂-Induced Apoptosis and Oxidative Stress in Human Lens Epithelial Cells via Inhibition of the NF- κ B Signaling Pathway. *Pharmacology.* 2018; 102: 213–222.
 86. Wei C, Li L, Kim IK, Sun P, Gupta S. NF- κ B mediated miR-21 regulation in cardiomyocytes apoptosis under oxidative stress. *Free Radic. Res.* 2014; 48: 282–291.
 87. Xie Y, Chu A, Feng Y, Chen L, Shao Y, et al. MicroRNA-146a: A comprehensive indicator of inflammation and oxidative stress status induced in the brain of chronic T2DM rats. *Front. Pharmacol.* 2018; 9: 478.
 88. Fariás JG, Molina VM, Carrasco RA, Zepeda AB, Figueroa E, et al. Antioxidant therapeutic strategies for cardiovascular conditions associated with oxidative stress. *Nutrients.* 2017; 9: 966.
 89. Sekhon MS, Ainslie PN, Griesdale DE. Clinical pathophysiology of hypoxic ischemic brain injury after cardiac arrest: A “two-hit” model. *Crit. Care.* 2017; 21: 90.
 90. Chekulaeva M, Filipowicz W. Mechanisms of miRNA-mediated post-transcriptional regulation in animal cells. *Curr. Opin. Cell Biol.* 2009; 21: 452–460.

91. Dong Y, Xu W, Liu C, Liu P, Li P, et al. Reactive oxygen species related noncoding RNAs as regulators of cardiovascular diseases. *Int. J. Biol. Sci.* 2019; 15: 680–687.
92. Takimoto E, Kass DA. Role of oxidative stress in cardiac hypertrophy and remodeling. *Hypertension.* 2007; 49: 241–248.
93. Zhao Y, Ransom JF, Li A, Vedantham V, von Drehle M, et al. Dysregulation of Cardiogenesis, Cardiac Conduction, and Cell Cycle in Mice Lacking miRNA-1-2. *Cell.* 2007; 129: 303–317.
94. Van Rooij E, Sutherland LB, Liu N, Williams AH, McAnally J, et al. A signature pattern of stress-responsive microRNAs that can evoke cardiac hypertrophy and heart failure. *Proc. Natl. Acad. Sci. USA.* 2006; 103: 18255–18260.
95. Ikeda S, He A, Kong SW, Lu J, Bejar R, et al. MicroRNA-1 Negatively Regulates Expression of the Hypertrophy-Associated Calmodulin and Mef2a Genes. *Mol. Cell. Biol.* 2009; 29: 2193–2204.
96. Carè A, Catalucci D, Felicetti F, Bonci D, Addario A, et al. MicroRNA-133 controls cardiac hypertrophy. *Nat. Med.* 2007; 13: 613–618.
97. Wojciechowska A, Braniewska A, Kozar-Kamińska K. MicroRNA in cardiovascular biology and disease. *Adv. Clin. Exp. Med.* 2017; 26: 865–874.
98. Van Rooij E, Sutherland LB, Qi X, Richardson JA, Hill J, et al. Control of stress-dependent cardiac growth and gene expression by a microRNA. *Science.* 2007; 316: 575–579.
99. Rawal S, Nagesh PT, Coffey S, Van Hout I, Galvin IF, et al. Early dysregulation of cardiac-specific microRNA-208a is linked to maladaptive cardiac remodelling in diabetic myocardium. *Cardiovasc. Diabetol.* 2019; 18: 13.
100. Fu J, Chen Y, Li F. Attenuation of MicroRNA-495 Derepressed PTEN to Effectively Protect Rat Cardiomyocytes from Hypertrophy. *Cardiology.* 2018; 139: 245–254.
101. Da Costa Martins PA, Salic K, Gladka MM, Armand AS, Leptidis S, et al. MicroRNA-199b targets the nuclear kinase Dyrk1a in an auto-amplification loop promoting

- calcineurin/NFAT signalling. *Nat. Cell Biol.* 2010; 12: 1220–1227.
102. Li Z, Song Y, Liu L, Hou N, An X, et al. MiR-199a impairs autophagy and induces cardiac hypertrophy through mTOR activation. *Cell Death Differ.* 2017; 24: 1205–1213.
103. Kura B, Parikh M, Slezak J, Pierce GN. The influence of diet on microRNAs that impact cardiovascular disease. *Molecules.* 2019; 24: 1509.
104. Dong D, Yang B. Role of microRNAs in cardiac hypertrophy, myocardial fibrosis and heart failure. *Acta Pharm. Sin. B* 2011; 1: 1–7.
105. Wang H, Cai J. The role of microRNAs in heart failure. *Biochim. Biophys. Acta.* 2017; 1863: 2019–2030.
106. Van Empel VPM, De Windt LJ, Da Costa Martins PA. Circulating miRNAs: Reflecting or affecting cardiovascular disease. *Curr. Hypertens. Rep.* 2012; 14: 498–509.
107. Sassi Y, Avramopoulos P, Ramanujam D, Grüter L, Werfel S, et al. Cardiac myocyte miR-29 promotes pathological remodeling of the heart by activating Wnt signaling. *Nat. Commun.* 2017; 8: 1614.
108. Piper HM, Meuter K, Schäfer C. Cellular mechanisms of ischemia-reperfusion injury. *Ann. Thorac. Surg.* 2003; 75: S644–S648.
109. Xiao X, Lu Z, Lin V, May A, Shaw DH, et al. MicroRNA miR-24-3p reduces apoptosis and regulates Keap1-Nrf2 pathway in mouse cardiomyocytes responding to ischemia/reperfusion injury. *Oxid. Med. Cell. Longev.* 2018; 2018: 7042105.
110. Jiang H, Lu Z. MicroRNA-144 attenuates cardiac ischemia/reperfusion injury by targeting FOXO1. *Exp. Ther. Med.* 2019; 17: 2152–2160.
111. Puthanveetil P, Zhang D, Wang Y, Wang F, Wan A, et al. Diabetes triggers a PARP1 mediated death pathway in the heart through participation of FoxO1. *J. Mol. Cell. Cardiol.* 2012; 53: 677–686.
112. Chen CJ, Yu W, Fu YC, Wang X, Li JL, et al. Resveratrol protects cardiomyocytes from hypoxia-induced apoptosis through the SIRT1-FoxO1 pathway. *Biochem. Biophys. Res. Commun.* 2009; 378: 389–393.

113. Fang YC, Yeh CH. Inhibition of MIR-302 Suppresses Hypoxia-Reoxygenation-Induced H9c2 Cardiomyocyte Death by Regulating Mcl-1 Expression. *Oxid. Med. Cell. Longev.* 2017; 2017: 7968905.
114. Woo CC, Wongsurawat T, Lin XY, Sorokin V. The miRNA 30B-5P targeting mRNA MBNL1 leads to pro-myogenic VSMC phenotype modulation in myocardial infarction patients. *Atherosclerosis.* 2018; 275: e46.
115. Woo CC, Wongsurawat T, Soong R, Lee CN, Richards M, et al. Distinctive pattern of LET-7B and MIR-30B in human aortic smooth muscle cells of myocardial infarction patients. *Atherosclerosis.* 2017; 263: e63.
116. Sorokin V, Woo CC, Lin XY, Kofidis T, Lee CN. Role of Micro RNA in Arterial Wall Remodeling in Patients with Advanced Coronary Artery Disease Undergoing Bypass. In *Proceedings of the 22nd Annual Meeting of the Asian Society for Cardiovascular and Thoracic Surgery (ASCVTS'14)*. Istanbul, Turkey. 2014.
117. Long B, Gan TY, Zhang RC, Zhang YH. miR-23a regulates cardiomyocyte apoptosis by targeting manganese superoxide dismutase. *Mol. Cells.* 2017; 40: 542–549.
118. Wang L, Yuan Y, Li J, Ren H, Cai Q, et al. MicroRNA-1 aggravates cardiac oxidative stress by post-transcriptional modification of the antioxidant network. *Cell Stress Chaperones.* 2015; 20: 411–420.
119. Li Y, Du Y, Cao J, Gao Q, Li H, et al. MiR-130a inhibition protects rat cardiac myocytes from hypoxia-triggered apoptosis by targeting Smad4. *Kardiol. Pol.* 2018; 76: 993–1001.
120. Sun C, Liu H, Guo J, Yu Y, Yang D, et al. MicroRNA-98 negatively regulates myocardial infarction-induced apoptosis by down-regulating Fas and caspase-3. *Sci. Rep.* 2017; 7: 7460.
121. Liu A, Sun Y, Yu B. Microrna-208a correlates apoptosis and oxidative stress induced by h2o2 through protein tyrosine kinase/phosphatase balance in cardiomyocytes. *Int. Heart J.* 2018; 59: 829–836.
122. Zhou L, Zang G, Zhang G, Wang H, Zhang X, et al. MicroRNA and mRNA signatures in ischemia reperfusion injury in heart transplantation. *PLoS ONE.* 2013; 8: e79805.

123. Shah P, Bristow MR, Port JD. MicroRNAs in Heart Failure, Cardiac Transplantation, and Myocardial Recovery: Biomarkers with Therapeutic Potential. *Curr. Heart Fail. Rep.* 2017; 14: 454–464.
124. Hamdorf M, Kawakita S, Everly M. The Potential of MicroRNAs as Novel Biomarkers for Transplant Rejection. *J. Immunol. Res.* 2017; 2017: 4072364.
125. Van Huyen JPD, Tible M, Gay A, Guillemain R, Aubert O, et al. MicroRNAs as non-invasive biomarkers of heart transplant rejection. *Eur. Heart J.* 2014; 35: 3194–3202.
126. Fang Y, Shi C, Manduchi E, Civelek M, Davies PF. MicroRNA-10a regulation of proinflammatory phenotype in athero-susceptible endothelium in vivo and in vitro. *Proc. Natl. Acad. Sci. USA.* 2010; 107: 13450–13455.
127. Suárez Y, Wang C, Manes TD, Pober JS. Cutting Edge: TNF-Induced MicroRNAs Regulate TNF-Induced Expression of E-Selectin and Intercellular Adhesion Molecule-1 on Human Endothelial Cells: Feedback Control of Inflammation. *J. Immunol.* 2010; 184: 21–25.
128. Zhang L, Zhou M, Wang Y, Huang W, Qin G, et al. MiR-92a inhibits vascular smooth muscle cell apoptosis: Role of the MKK4-JNK pathway. *Apoptosis.* 2014; 19: 975–983.
129. Wei L, Wang M, Qu X, Mah A, Xiong X, et al. Differential expression of microRNAs during allograft rejection. *Am. J. Transplant.* 2012; 12: 1113–1123.
130. Malakar AK, Choudhury D, Halder B, Paul P, Uddin A, et al. A review on coronary artery disease, its risk factors, and therapeutics. *J. Cell. Physiol.* 2019; 234: 16812–16823.
131. Li T, Song X, Zhang J, Zhao L, Shi Y, et al. Protection of Human Umbilical Vein Endothelial Cells against Oxidative Stress by MicroRNA-210. *Oxid. Med. Cell. Longev.* 2017; 2017: 3565613.
132. Fu XM, Zhou YZ, Cheng Z, Liao XB, Zhou XM. MicroRNAs: Novel players in aortic aneurysm. *Biomed. Res. Int.* 2015; 2015: 831641.
133. Marcil V, Delvin E, Sané AT, Tremblay A, Levy E. Oxidative stress influences cholesterol efflux in THP-1 macrophages: Role of ATP-binding cassette A1 and nuclear factors. *Cardiovasc. Res.* 2006; 72: 473–482.

134. Kataoka M, Wang DZ. Non-Coding RNAs Including miRNAs and lncRNAs in Cardiovascular Biology and Disease. *Cells*. 2014; 3: 883–898.
135. Qu X, Du Y, Shu Y, Gao M, Sun F, et al. MIAT is a Pro-fibrotic Long Non-coding RNA Governing Cardiac Fibrosis in Post-infarct Myocardium. *Sci. Rep.* 2017; 7: 42657.
136. Zheng Y, Li Y, Liu G, Qi X, Cao X. MicroRNA-24 inhibits the proliferation and migration of endothelial cells in patients with atherosclerosis by targeting importin- α 3 and regulating inflammatory responses. *Exp. Ther. Med.* 2018; 15: 338–344.
137. Zhang J, Cai W, Fan Z, Yang C, Wang W, et al. MicroRNA-24 inhibits the oxidative stress induced by vascular injury by activating the Nrf2/Ho-1 signaling pathway. *Atherosclerosis*. 2019; 290: 9–18.
138. Gou L, Zhao L, Song W, Wang L, Liu J, et al. Inhibition of miR-92a Suppresses Oxidative Stress and Improves Endothelial Function by Upregulating Heme Oxygenase-1 in db/db Mice. *Antioxid. Redox Signal.* 2018; 28: 358–370.
139. Yamac AH, Huyut MA, Yilmaz E, Celikkale I, Bacaksiz A, et al. MicroRNA 199a is downregulated in patients after coronary artery bypass graft surgery and is associated with increased levels of sirtuin 1 (SIRT 1) protein and major adverse cardiovascular events at 3-year follow-up. *Med. Sci. Monit.* 2018; 24: 6245–6254.
140. Guo M, Mao X, Ji Q, Lang M, Li S, et al. MiR-146a in PBMCs modulates Th1 function in patients with acute coronary syndrome. *Immunol. Cell Biol.* 2010; 88: 555–564.
141. O’Sullivan JF, Neylon A, McCorrigan C, Blake GJ. miRNA-93-5p and other miRNAs as predictors of coronary artery disease and STEMI. *Int. J. Cardiol.* 2016; 224: 310–316.
142. Tsutsui H, Kinugawa S, Matsushima S. Oxidative stress and heart failure. *Am. J. Physiol. Heart. Circ. Physiol.* 2011; 301: H2181–H2190.
143. Dickstein K, Cohen-Solal A, Filippatos G, McMurray JJV, Ponikowski P, et al. ESC Guidelines for the diagnosis and treatment of acute and chronic heart failure 2008. The Task Force for the Diagnosis and Treatment of Acute and

- Chronic Heart Failure 2008 of the European Society of Cardiology. Developed in collaboration with the Heart. *Eur. J. Heart Fail.* 2008; 10: 933–989.
144. Hunt SA, Abraham WT, Chin MH, Feldman AM, Francis GS, et al. 2009 Focused Update Incorporated Into the ACC/AHA 2005 Guidelines for the Diagnosis and Management of Heart Failure in Adults: A Report of the American College of Cardiology Foundation/American Heart Association Task Force on Practice Guidelines Developed. *J. Am. Coll. Cardiol.* 2009; 53: e1–e90.
145. Hamaguchi S, Kinugawa S, Tsuchihashi-Makaya M, Goto K, Goto D, et al. Spironolactone use at discharge was associated with improved survival in hospitalized patients with systolic heart failure. *Am. Heart J.* 2010; 160: 1156–1162.
146. Hamaguchi S, Tsuchihashi-Makaya M, Kinugawa S, Yokota T, Ide T, et al. Chronic kidney disease as an independent risk for long-term adverse outcomes in patients hospitalized with heart failure in Japan—Report from the Japanese Cardiac Registry of Heart Failure in Cardiology (JCARE-CARD). *Circ. J.* 2009; 73: 1442–1447.
147. Tsutsui H, Tsuchihashi-Makaya M, Kinugawa S, Goto D, Takeshita A. Clinical characteristics and outcome of hospitalized patients with heart failure in Japan—Rationale and design of Japanese Cardiac Registry Of Heart Failure In Cardiology (JCARE-CARD). *Circ. J.* 2006; 70: 1617–1623.
148. Tsuchihashi-Makaya M, Hamaguchi S, Kinugawa S, Yokota T, Goto D, et al. Characteristics and outcomes of hospitalized patients with heart failure and reduced vs preserved ejection fraction—A report from the Japanese Cardiac Registry of Heart Failure in Cardiology (JCARE-CARD). *Circ. J.* 2009; 73: 1893–1900.
149. Grieve DJ, Shah AM. Oxidative stress in heart failure. More than just damage. *Eur. Heart J.* 2003; 24: 2161–2163.
150. Sawyer DB. Oxidative stress in heart failure: What are we missing? *Am. J. Med. Sci.* 2011; 342: 120–124.
151. van der Pol A, van Gilst WH, Voors AA, van der Meer P. Treating oxidative stress in heart failure: Past, present and future. *Eur. J. Heart Fail.* 2019; 21: 425–435.

152. Shan H, Zhang Y, Lu Y, Zhang Y, Pan Z, et al. Downregulation of miR-133 and miR-590 contributes to nicotine-induced atrial remodelling in canines. *Cardiovasc. Res.* 2009; 83: 465–472.
153. Ye Y, Perez-polo JR, Qian J, Birnbaum Y, Ye Y, et al. The role of microRNA in modulating myocardial ischemia-reperfusion injury The role of microRNA in modulating myocardial ischemia-reperfusion injury. *Physiol. Genom.* 2010; 43: 534–542.
154. Zhang X, Price NL, Fernández-Hernando C. Non-coding RNAs in lipid metabolism. *Vascul. Pharmacol.* 2019; 114: 93–102.
155. Schulte C, Karakas M, Zeller T. MicroRNAs in cardiovascular disease—Clinical application. *Clin. Chem. Lab. Med.* 2017; 55: 687–704.
156. Ali SS, Kala C, Abid M, Ahmad N, Sharma US, et al. Pathological microRNAs in acute cardiovascular diseases and microRNA therapeutics. *J. Acute Dis.* 2016; 5: 9–15.
157. Lin H, Sue YM, Chou Y, Cheng CF, Chang CC, et al. Activation of a nuclear factor of activated T-lymphocyte-3 (NFAT3) by oxidative stress in carboplatin-mediated renal apoptosis. *Br. J. Pharmacol.* 2010; 161: 1661–1676.
158. Kakita T, Hasegawa K, Iwai-Kanai E, Adachi S, Morimoto T, et al. Calcineurin pathway is required for endothelin-1-mediated protection against oxidant stress-induced apoptosis in cardiac myocytes. *Circ. Res.* 2001; 88: 1239–1246.
159. Wang X, Lian Y, Wen X, Guo J, Wang Z, et al. Expression of miR-126 and its potential function in coronary artery disease. *Afr. Health Sci.* 2017; 17: 474–480.
160. Reddy LL, Shah SAV, Ponde CK, Rajani RM, Ashavaid TF. Circulating miRNA-33: A potential biomarker in patients with coronary artery disease. *Biomarkers.* 2019; 24: 36–42.
161. Schulte C, Zeller T. microRNA-based diagnostics and therapy in cardiovascular disease-Summing up the facts. *Cardiovasc. Diagn. Ther.* 2015; 5: 17–36.
162. Cakmak HA, Barman HA, Coskunpinar E, Oltulu YM, Ikitimur B, et al. The Diagnostic Importance of MicroRNAs

- in Congestive Heart Failure. *J. Am. Coll. Cardiol.* 2013; 62: C17–C18.
163. Murach KA, McCarthy JJ. MicroRNAs, heart failure, and aging: Potential interactions with skeletal muscle. *Heart Fail. Rev.* 2017; 22: 209–218.
164. Lind EF, Ohashi PS. Mir-155, a central modulator of T-cell responses. *Eur. J. Immunol.* 2014; 44: 11–15.
165. Jones Buie JN, Goodwin AJ, Cook JA, Halushka PV, Fan H. The role of miRNAs in cardiovascular disease risk factors. *Atherosclerosis.* 2016; 254: 271–281.
166. Gilad S, Meiri E, Yogeve Y, Benjamin S, Lebanony D, et al. Serum microRNAs are promising novel biomarkers. *PLoS ONE.* 2008; 3: e3148.
167. Siebolts U, Vamholt H, Drebber U, Dienes HP, Wickenhauser C, et al. Tissues from routine pathology archives are suitable for microRNA analyses by quantitative PCR. *J. Clin. Pathol.* 2009; 62: 84–88.
168. Gaarz A, Debey-Pascher S, Classen S, Eggle D, Gathof B, et al. Bead array-based microRNA expression profiling of peripheral blood and the impact of different RNA isolation approaches. *J. Mol. Diagnostics.* 2010; 12: 335–344.
169. Miranda KC, Huynh T, Tay Y, Ang YS, Tam WL, et al. A Pattern-Based Method for the Identification of MicroRNA Binding Sites and Their Corresponding Heteroduplexes. *Cell.* 2006; 126: 1203–1217.
170. Olson EN. MicroRNAs as therapeutic targets and biomarkers of cardiovascular disease. *Sci. Transl. Med.* 2014; 6: 239ps3.
171. Wang HR, Wu M, Yu H, Long S, Stevens A, et al. Selective inhibition of the Kir2 family of inward rectifier potassium channels by a small molecule probe: The discovery, SAR, and pharmacological characterization of ML133. *ACS Chem. Biol.* 2011; 6: 845–856.
172. Krützfeldt J, Rajewsky N, Braich R, Rajeev KG, Tuschl T, et al. Silencing of microRNAs in vivo with antagomirs. *Nature.* 2005; 438: 685–689.
173. Oliveira-Carvalho V, Carvalho VO, Silva MM, Guimarães GV, Bocchi EA. MicroRNAs: A new paradigm in the treatment and diagnosis of heart failure? *Arq. Bras. Cardiol.* 2012; 98: 362–369.

174. Ebert MS, Neilson JR, Sharp PA. MicroRNA sponges: Competitive inhibitors of small RNAs in mammalian cells. *Nat. Methods*. 2007; 4: 721–726.
175. Ebert MS, Sharp PA. MicroRNA sponges: Progress and possibilities. *RNA*. 2010; 16: 2043–2050.
176. Broderick JA, Zamore PD. MicroRNA therapeutics. *Gene Ther*. 2011; 18: 1104–1110.
177. Caroli A, Cardillo MT, Galea R, Biasucci LM. Potential therapeutic role of microRNAs in ischemic heart disease. *J. Cardiol*. 2013; 61: 315–320.