

Book Chapter

Reduction of Serotonergic Gene Expression in the Midbrain Raphe Nuclei under Positive Fighting Experience

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Abstract

The concept of a major inhibitory role of serotonin in aggressive behavior is widely accepted by investigators. There was ample evidence that a pharmacologically-induced increase in the serotonin activity attenuates agonistic behavior in animals and that the manipulations inhibiting the brain serotonergic system can elicit aggressiveness in male mice and rats. Repeated experience of aggression in daily agonistic interactions has been shown to reduce serotonin activity in brain of victorious male mice. The study aimed to analyze expression of the serotonergic genes - *Tph2*, *Sert*, *Maoa* and *5ht1a*, as well as *Bdnf* and *Creb* genes in the midbrain raphe nuclei of male mice with positive fighting experience in daily encounters and male mice with the same track record of aggression followed by two-week no-fight period. It has been shown that mRNA levels of the serotonergic and *Creb* genes are reduced in the winners in comparison with male mice without consecutive positive fighting experience of aggression. After the fighting deprivation the *Tph2*, *Sert*, *Bdnf* and *Creb* genes recover their expression while mRNA levels of *Maoa* and *5ht1a* gene proceed at a significantly higher level as compared with the respective controls. Downregulation of serotonergic genes is indicative of the inhibition of serotonergic activity under repeated experience of aggression. Nevertheless, recovering of *Tph2* and *Sert* gene expression and overexpression of *Maoa* and *5ht1a* genes after no-fight period suggest that changes in brain serotonergic activity are not main cause of the behavioral pathology developing in male mice in this experimental context.

Keywords

Tph2; *Sert*; *Maoa*; *Bdnf*; *Creb*; mRNA; Repeated Aggression; Fighting Deprivation

Introduction

The concept of a major inhibitory role of serotonin (5-HT) in aggressive behavior is widely accepted by investigators. There was ample evidence that a pharmacologically-induced increase in 5-HT activity attenuates agonistic behavior in animals and that the manipulations inhibiting the brain serotonergic system can elicit aggressiveness in male mice and rats [1-7]. It has been shown that after fights 5-HT level decreases in the prefrontal cortex in aggressive rats [8]. Reduced cerebrospinal fluid (CSF) level of the 5-HT major metabolite 5-hydroxyindoleacetic acid (5-HIAA), as well as decreased sensitivity of 5-HT_{1A} receptors shown in prisoners or psychiatric patients with repeated violent or aggressive acts [9-13], also suggests a reduced overall 5-HT function in aggressive individuals.

Experimental studies in male mice with repeated experience of aggression accompanied by social victories in daily agonistic interactions have demonstrated reduced 5-HIAA level or/and 5-HIAA/5-HT ratio [14-15] and a decrease in the activity of rate-limiting enzyme in the 5-HT biosynthesis, tryptophan hydroxylase, in brain areas such as midbrain, hippocampus and striatum [16,17]. Specific changes in pharmacological sensitivity of 5-HT_{1A} receptors to agonist of 5-HT_{1A} receptor buspirone [18] and in 5-HT_{1A} receptor binding in the frontal cortex, hippocampus and hypothalamus have been demonstrated in male mice with repeated aggression [19]. All these findings are consistent with the 5-HT deficiency hypothesis of aggression [15].

The study aimed to elicit a possible role of serotonergic *Tph2*, *Sert*, *Maoa*, *5ht1a* genes, encoding tryptophan hydroxylase, the 5-HT transporter, monoamine oxidase A and 5HT_{1A} receptors, respectively - may be involved into regulations of positive fighting experience. Expression of the *Bdnf* (encoding brain-

derived neurotrophic factor) and *Creb* (encoding cyclic AMP response element binding protein) genes were also studied. The focus was on the area of the midbrain raphe nuclei containing the cell bodies of serotonergic neurons. The mRNA levels were analyzed in male mice that had a long positive fighting history (21 wins in daily agonistic interactions). Since the behavioral data indicated that the implications of chronic aggression persist at least two weeks after the cessation of agonistic interactions [15,20,21], the expression of serotonergic genes was also analyzed in a group of 21-time winners who were kept away from agonistic interactions referred to as "no-fight period" or "fighting deprivation" throughout. Such animals are special as they appear to be more aggressive after no-fight period than before. The comparison of the expression of genes in these groups of the winners helps to answer the question on whether or not the altered gene expression in the midbrain raphe nuclei of the fighting deprived winners recovers after no-fight period to the control levels.

Materials and Methods

Animals

Adult male mice of the C57BL/6J strain from a stock maintained in the Animal Facility of the Institute of Cytology and Genetics, SD RAS, (Novosibirsk, Russia) were used. The animals were housed under standard conditions [12:12-h light/dark regime, switch on at 8.00 a.m.; food (pellets) and water available *ad libitum*]. Mice were weaned at one month of age and housed in groups of eight to ten in plastic cages (36 x 23 x 12 cm). Experiments were performed on mice 10-12 weeks of age.

All procedures were in compliance with the European Communities Council Directive of November 24, 1986 (86/609/EEC). The study protocol was approved by the Scientific Committee N 9 on the Ethics of Animal Experiments of the Institute of Cytology and Genetics SD RAS (Permit Number: N 613).

Sensory Contact Model for the Studying Agonistic Behavior in Male Mice

Repeated experience of aggression accompanied by social victories in male mice was induced using the sensory contact model [22-24]. Pairs of weight-matched animals were each placed in a steel cage bisected by a perforated transparent partition allowing the animals to see, hear and smell each other, but preventing physical contact. The animals were left undisturbed for three days to adapt to new housing conditions and sensory contact before they were exposed to encounters. Every afternoon (1400-1700 hours, local time), the cage lid was replaced by a transparent one, and 5 min later (the period necessary for individuals' activation), the partition was removed for 10 minutes to encourage agonistic interactions. The superiority of one of the mice was firmly established within two or three encounters with the same opponent. The superior mouse (winners) would be attacking, biting and chasing another, who would be displaying only defensive behavior (sideways postures, upright postures, withdrawal, lying on the back or freezing). As a rule, agonistic interactions between males are discontinued by lowering the partition if the strong aggression has lasted 3 min, in some cases, less. Each defeated mouse (defeater, loser) was exposed to the same winner for three days, while afterwards each loser was placed, once a day after the fight, in an unfamiliar cage with an unfamiliar winner behind the partition. Each victorious mouse remained in its original cage. This procedure was performed once a day for 21 days and yielded an equal number of the winners and losers.

Three groups of animals were used. (1) Winners – a group of mice with repeated experience of aggression accompanied by victories during 21 days of agonistic interactions; (2) Fighting deprived winners - a group of chronically victorious 21-time mice who lived for 14 days after the last encounter without agonistic interactions (period of fighting deprivation). During this period, each of them shared a cage with losers; the partition between their compartments being down at all times, to prevent physical encounters. (3) Controls: the mice without consecutive experience of agonistic interactions. To the end of experiment,

animals of all experimental groups were 15-17 weeks of age.

To measure mRNA levels in the midbrain raphe nuclei area, all the mice were decapitated simultaneously: 21-time winners, 24 hours after the last agonistic interaction; fighting deprived winners, immediately after 14-day period of deprivation; and the controls. The raphe nuclei of midbrain were dissected according to the Mouse Brain Atlas [25] in animals of all experimental groups. The mouse brains were removed and chilled rapidly on ice. Dissection of the brain regions was made by one experimenter during two consecutive days. Animals of all groups were represented in each day. All biological samples were encrypted, rapidly frozen in liquid nitrogen, and stored at -70° C until use.

Behavioral Study

Behavior of each winner in agonistic interaction test was video recorded for 10 min during its last encounter, and the data were documented. Furthermore, we needed to know whether both groups of winners could be considered identical at the time each mouse was won its last encounter. To find out, the groups were compared in terms of behavior. During a 10-min test, the following were the behavioral domains analyzed in the winners: (1) *Attacks*: attacking, biting and chasing; (2) *Aggressive grooming* – the winner mounts onto the loser's back, holds it down and spends much time licking and nibbling at the loser's scruff of the neck. The loser is wholly immobilized or sometimes stretches out the neck and again freezes under the winner; (3) *Digging* – digging up and scattering the sawdust on the loser's territory (kick-digs: pulling the sawdust forwards with the forepaws; push-digs: pushing the sawdust backwards with the hind paws); (4) *Hostile behavior* – the total time spent attacking, aggressive grooming and digging; (5) *Self-grooming* - body care activities (fur licking, head washing, nose washing). The total time and/or number of behavioral events were measured.

Total RNA Extraction and Reverse Transcription

Total RNA was isolated from the tissues using TRIzol (Invitrogen) according to the manufacturer's instructions. The concentration of total RNA was quantified by measuring the absorbance at 260 nm. The integrity of total RNA was assessed using agarose gel electrophoresis. cDNAs were synthesized using total RNA (1 µg), random N₉ primer (100 ng), and MoMLV reverse transcriptase (200 U, Biosan). Each RT reaction was run in duplicate. RNA aliquots were used to confirm the absence of any genomic DNA in each sample.

Real-Time Quantitative PCR

Tph2, *Sert (Slc6a4)*, *Maoa*, *5ht1a (Htr1a)*, *Bdnf*, *Creb*, *Cphn*, *Gapdh*, and *Hprt* cDNA levels were quantified by SybrGreen-based real-time PCR in a total volume of 25 µl containing an aliquot of the RT mixture, dNTPs (200 nM), F and R primers (300 nM), SybrGreen I (1:20,000, Invitrogen), standard PCR buffer, and hot-start TaqDNA polymerase (0.5 U, Biosan). Amplification was run for 3 min at 95°C followed by 40 cycles of 6 s at 96°C, 6 s at 60°C, 12 s at 72°C. Fluorescence was monitored for 5 s after each cycle at the appropriate melting temperature. To check for the presence of nonspecific PCR products or primer dimers, a melting curve analysis was performed after the final PCR cycle. *Cphn*, *Gapdh*, and *Hprt* were considered initially as possible housekeeping (normalizers) genes in this study.

Amplification efficiencies were calculated using a relative standard curve derived from threefold serial dilutions of pooled cDNA. In all cases, the amplification efficiency was higher than 90%. Each sample was PCR-amplified twice. qRT-PCR results were quantified using the relative standard curve method. The PCR primer sequences are shown in Table 1. Samples from animals of different experimental groups were mixed before molecular study. Extraction of total RNA, reverse transcription, and qRT-PCR were made during 2 weeks by one experimenter in blind regimen with use of one buffer and primers set.

Table 1: Primer Sequences.

Genes	Primer sequences		Functions
<i>Bdnf</i>	F	5'-CAAACAAGACACATTACCTTCCT-3'	Brain-derived neurotrophic factor
	R	5'-ATGGTCATCACTCTTCTCACCT-3'	
<i>Creb</i>	F	5'-CAGCCACAGATTGCCACATTAG-3'	Cyclic AMP response element binding protein
	R	5'-CTTATGGAGACTGGATAACTGATG-3'	
<i>Tph2</i>	F	5'-CACCATTGTGACCCTGAATCC-3'	Tryptophan hydroxylase 2 - rate-limiting enzyme of 5-HT synthesis
	R	5'-AAGCTCGGTGCCGTACATGAG-3'	
<i>Sert</i>	F	5'-GCTGAGATGAGGAACGAAGAC-3'	Serotonin transporter protein
	R	5'-AGGAAGAAGATGATGGCAAAG-3'	
<i>Maoa</i>	F	5'-GAATGTCAATGAGCGTCTAGTTC-3'	Monoamine oxidase A, enzyme of 5-HT catabolism
	R	5'-ATGGTGCATCAACAGGGATCTC-3'	
<i>5ht1a</i>	F	5'-TTGGAACACTTTGGGTTATGG-3'	Serotonin 5HT1A receptors
	R	5'-ATTGTCAATTTCTTTGGTGAGTG-3'	
<i>Cphn</i>	F	5'-GTTTTTTATCTGCACTGCCAAG-3'	PPIA - peptidylprolyl isomerase A (cyclophilin A); enzyme accelerates folding of proteins
	R	5'-TTCTTGCTGGTCTTGCCATTC-3'	
<i>Gapdh</i>	F	5'-TGTTCCAGTATGACTCCACTCA-3'	Glyceraldehyde-3-phosphate dehydrogenase; glycolysis enzyme
	R	5'-GACACCAGTAGACTCCACGACA-3'	
<i>Hprt</i>	F	5'-TGAAAAGGACCTCTCGAAGTGT-3'	Hypoxanthine-guanine phosphoribosyltransferase; synthesis of purine nucleotides through the purine salvage pathway
	R	5'-CACTAATGACACAAACGTGATTC-3'	

Statistical Analysis

Normal distribution and homogeneity of variances were tested by the Shapiro–Wilk’s and Levene’s tests, respectively. Statistical analysis of behavioral data was performed pairwise comparison of the groups with t-Student test or the Mann-Whitney *U* test. Statistical analysis of mRNA levels was performed using one-way ANOVA with factor “groups” under the consideration — the controls, winners, and fighting deprived winners — followed by the comparison of the groups using the Bonferroni *post hoc* testing or Fisher LSD test. The data are reported as mean \pm SEM. The statistical significance was set at $P \leq 0.05$.

For each experimental sample, the relative amount of mRNA is determined from the appropriate standard curve. Since the all reference genes, *Cphn*, *Gapdh*, and *Hprt*, changed their expression under repeated experience of aggression in the midbrain raphe nuclei in the winners as compared with the controls, we can’t use them to obtain a normalized level of the *Bdnf*, *Creb*, *Tph2*, *Sert*, *Maoa*, and *5ht1a* genes. The other statistical approach [26] was used to reveal differences in serotonergic and other genes expression between the experimental groups: the mean of relative amount of mRNA in the control group for each gene was taken as 100%, and changes in the winners, fighting deprived winners, and controls were calculated as percentage of the mean in the controls. In the control group, the number of animals used in experiment was 16–17. Groups of 21-time winners and fighting deprived winners contained 11-13 mice.

Results

Analysis of the Winners’ Behavior in Agonistic Interactions

We needed at first to know whether both groups of the aggressive mice (winners) could be considered identical at the time each mouse was won its last encounter. If they were, all the differences in genes’ expression between group of 21-time winners and group of 21-time winners before fighting

deprivation, or lack thereof, could solely be accounted for by deprivation. To find out, the animals were compared in terms of aggressive behavior: number and/or total time of attacks, aggressive grooming, digging etc. No differences were found between these groups of the winners in any of the individual or social behaviors measured after the respective 21-day periods of agonistic interactions ($P > 0.05$, Table 2).

Table 2: Behavioral data from mice to become no-deprivation winners and in mice to become post-deprivation winners during their respective last encounter.

Behavioral parameters	Mice to become no-deprived winners	Mice to become post-deprived winners	Unpaired t-test M-W test
Attacks			
Latency, s	54,7 ± 15,7	55,4 ± 17,8	t = 0.030; $P < 0.97$
Number	11,2 ± 1,9	9,0 ± 1,7	t = 0.848; $P < 0.40$,
Total time, s	50,5 ± 12,1	43,7 ± 7,8	t = 0.848; $P < 0.64$,
Aggressive grooming, s	27,4 ± 17,2	23,4 ± 16,9	U=76; $P < 0.98$
Diggings, s	18,1 ± 3,5	18,6 ± 2,4	t = 0.126; $P = 0.90$,
Hostile behavior, s	95,9 ± 18,8	85,7 ± 17,8	t = 0.386; $P < 0.70$
Self-grooming, N	19,2 ± 3,7	14,9 ± 3,6	t = 0.817; $P = 0.42$
Number of animals	14	11	

Cphn, *Gapdh*, and *Hprt* Genes

Data obtained revealed a change in expression of the so-called housekeeping genes which we were going to use for normalization for the qRT PCR data: the expression of the *Cphn*, *Gapdh*, and *Hprt* genes used as putative reference genes decreased significantly in the midbrain raphe nuclei in the 21-time winners as compared with the controls. Similarly to previous study [26] as it turns out, the use of these genes as reference genes in the analysis of expression of other genes of interest was hardly possible. The method for calculating changes in the expression of each gene in the winners before and after deprivation versus its expression in the respective controls was

used to reveal differences in all genes expression between the experimental groups [26].

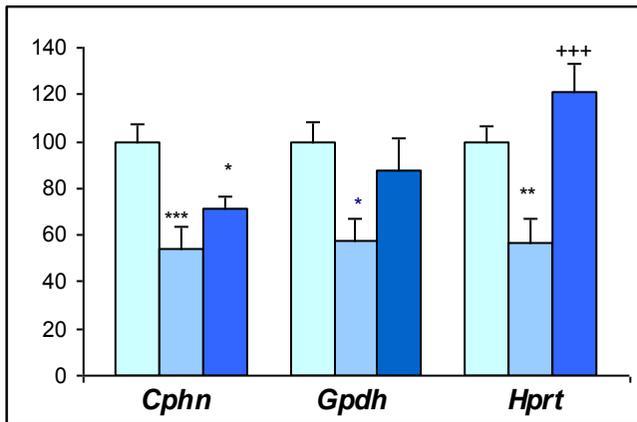


Figure 1: *Cphn*, *Gapdh*, and *Hprt* mRNA levels in the raphe nuclei of the midbrain in the controls (light columns), winners (grey columns), and fighting deprived winners (dark columns). Data are presented as percentage of the mean in the controls. * - $P < 0.05$; ** - $P < 0.01$; *** - $P < 0.001$ vs the controls; and +++ - $P < 0.001$ vs the winners.

One-way ANOVA revealed a significant influence of the factor groups (the control, winners and fighting deprived winners) on mRNA level of the *Cphn* ($F_{2,38} = 9.58$; $P < 0.001$), *Gapdh* ($F_{2,37} = 4.77$; $P < 0.014$), and *Hprt* ($F_{2,39} = 10.50$; $P < 0.001$) genes (Fig. 1). Based on the *post hoc* Bonferroni test as compared to the respective levels in the controls, mRNA levels of the *Cphn*, *Gapdh*, and *Hprt* genes were decreased in the 21-time winners ($P < 0.001$, $P < 0.013$, $P < 0.007$, respectively). In fighting deprived winners *Cphn* mRNA level was significantly less ($P < 0.035$) and expression of the *Gapdh* and *Hprt* genes did not differ significantly in comparison with the controls. However, expression of the *Hprt* gene in the fighting deprived winners was significantly higher than in the winners before deprivation ($P < 0.001$).

Bdnf and *Creb* Genes

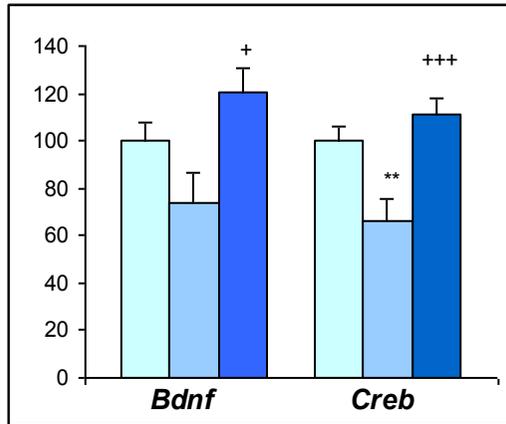


Figure 2: *Bdnf* and *Creb* mRNA levels in the raphe nuclei of the midbrain controls (light columns), winners (grey columns), and fighting deprived winners (dark columns). Data are presented as percentage of the mean in the controls. ** - $P < 0.01$; vs the controls; + - $P < 0.05$; +++ - $P < 0.001$ vs the winners.

ANOVA revealed a significant influence of the factor groups on mRNA levels of the *Bdnf* ($F_{2,39} = 4.76$; $P < 0.014$) and *Creb* ($F_{2,38} = 8.88$; $P < 0.001$) genes. Based on the Bonferroni *post hoc* test (Fig. 2) as compared to the respective levels in the controls, mRNA levels of the *Creb* gene were decreased ($P < 0.008$) in the 21-time winners. In the fighting deprived winners, *Bdnf* and *Creb* mRNA level was higher than in the winners before deprivation ($P < 0.012$ and $P < 0.001$, respectively).

Tph2, *Sert*, *Maoa*, and *5ht1a* Genes

One-way ANOVA revealed a significant influence of the factor groups on mRNA level of the *Tph2* ($F_{2,36} = 5.36$; $P < 0.009$), *Sert* ($F_{2,38} = 4.79$; $P < 0.014$), *Maoa*, ($F_{2,39} = 15.32$; $P < 0.001$) and *5ht1a* ($F_{2,38} = 37.84$; $P < 0.001$) genes. Based on the Bonferroni *post hoc* test (Fig. 3) as compared to the respective controls, mRNA levels of the *Maoa* and *5ht1a* genes were decreased in the 21-time winners ($P < 0.007$ and $P < 0.037$, respectively) and was increased in fighting deprived winners (P

< 0.033 and $P < 0.001$, respectively). Significant differences were found between the winners before and after deprivation for the *Tph2* ($P < 0.007$), *Sert* ($P < 0.012$), *Maoa* ($P < 0.001$), and *5ht1a* ($P < 0.001$) genes. Additionally, Fisher LSD test had demonstrated significant differences between the controls and winners for *Sert* mRNA levels ($P < 0.043$).

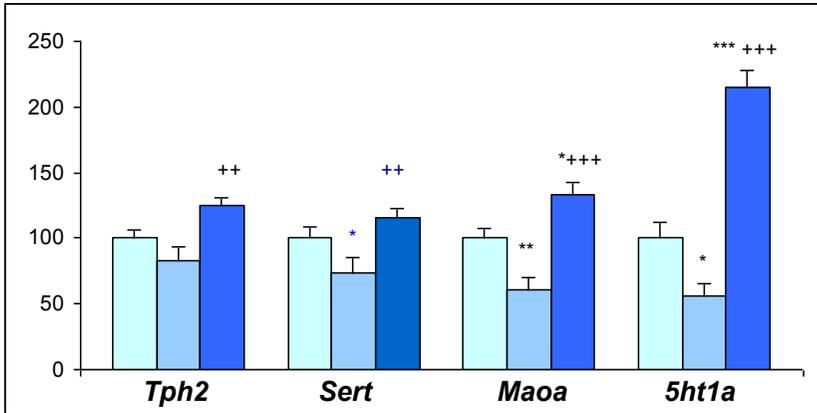


Figure 3: *Tph2*, *Sert*, *Maoa*, and *5ht1a* mRNA levels in the raphe nuclei of the midbrain in the controls (light columns), winners (grey columns), and fighting deprived winners (dark columns). Data are presented as % of the mean in the controls. * - $P < 0.05$; ** - $P < 0.01$ and *** - $P < 0.01$ vs the controls; ++ - $P < 0.01$; +++ - $P < 0.001$ vs the winners.

Discussion

Variability of reference genes expression in different tissues and experimental situations was demonstrated in studies of last years [27-32]. We also found in the 21 time winners decrease in functional activity of the genes involved in the processes of intracellular protein transport (*Cphn*) and glycolysis (*Gapdh*), presented in all nucleated cells (*Hprt*) that suggests a profound influence of positive fighting experience on different metabolic and cellular processes in the raphe nuclei of the midbrain. It is noteworthy that 2 week no-fight period without agonistic interactions is enough for *Gapdh* and *Hprt* genes but not for *Cphn* gene to recover their expression to the control levels.

Transcription of *Bdnf* is dependent on cAMP response element binding protein (CREB). In the midbrain raphe nuclei of the winners before and after fighting deprivations the expression of the *Bdnf* and *Creb* genes, encoding the proteins associated with neurotransmission, neurogenesis and synaptic plasticity [33] was shown to undergo changes similar to those in genes involved into the metabolic and cellular processes. Reduced *Creb* mRNA level was found in the midbrain raphe nuclei of 21-time winners as compared with the controls. It is notable that mRNA levels of both genes were significantly higher after a no-fight period than before it. Dynamic changes in the expression of these genes are likely to take place under experience of aggression and no-fight periods.

It has been shown many times that the *Bdnf* and *Creb* genes are involved in the brain mechanism of chronic social defeat stress [26,34-38]. At the same time, the experimental evidence of the role of these genes in the mechanisms of aggression is scarce and inconclusive. It has been shown that BDNF knockout mice exhibit elevated conspecific aggression and social dominance [39]. Schizophrenic patients with first-episode psychosis have lower serum concentrations of BDNF and score higher on the Aggression-Hostility scale than healthy individuals [40]. Winning animals have significantly more *Bdnf* mRNA in the dentate gyrus of the dorsal hippocampus than losing animals and home cage controls [41]. Authors support a model in which BDNF-mediated plasticity within the hippocampus may instantiate aspects of winning such as control of a territory by dominant animals. *Bdnf* mRNA level is increased in ventral tegmental area of 21-time winners [42]. Obviously, further studies are needed to explore the role of the *Bdnf* and *Creb* in the mechanisms of social aggression.

Research evidence shows that BDNF promotes the survival and differentiation of 5-HT neurons [43]. It is reasonable to assume that changes in the functional activity of the *Creb* and *Bdnf* genes in the midbrain raphe nuclei containing the cell bodies of serotonergic neurons are attributable to changes in serotonergic activity of the brain shaped by repeated experience of aggression. The experiment demonstrated a significantly reduced

expression of the *Maoa*, *5ht1a*, *Sert*, and lesser reduction - *Tph2* mRNA levels in the mice winning 21 successive encounters each. It is noteworthy that earlier a decrease in the activity of rate-limiting enzyme in the biosynthesis of 5-HT, tryptophan hydroxylase in the midbrain was shown for the winners [16,17]. Thus, a chronic manifestation of aggression, which, as supposed earlier, is accompanied by the inhibition of the brain serotonergic system [15], decreases the expression of the serotonergic genes, whose products are responsible for the inactivation and reception of 5-HT. After no-fight period expression of the *Tph2* and *Sert* genes increases significantly versus the period before deprivation and to a larger degree for *Maoa*, and *5ht1a* genes versus the respective controls. Overexpression of genes after deprivation may be considered as common feedback mechanism initiated by cessation of agonistic interactions. The data obtained so far strongly support the statement that the serotonergic genes are involved in the mechanisms of repeated aggression and do not contradict the serotonin deficiency hypothesis of aggression [4,6], which associates the decreased 5-HT function and increased aggression. From this statement, it follows that serotonin inhibits aggression. Based on our data it is plausible to assume that the repeated experience of aggression inhibits the brain serotonergic activity and changes the 5-HT metabolism, reception and serotonergic gene expression.

It is noteworthy, that reduced mRNA levels of genes of interest in this area have been found in the animals with repeated experience of defeats [26]. Similarity of changes in male mice with alternative social behaviors, manifested by chronic defeats or aggression cast doubt on the specificity of such changes for aggression. Earlier we assumed [44] that similar changes in the winners and losers may be explained by influence of agonistic interactions stress common for both participants of social conflicts or by development of enhanced level of anxiety shown both in the winners [24] and losers [45]. However, we should also take into consideration the different dynamics of gene expression changes during period without agonistic interactions. In the losers, period of 2 weeks of relative rest is not enough to recover expression of all serotonergic genes as well as the *Creb* gene to the control levels [26]. In the winners during no-fight

period most genes fully restore their expression. The *Maoa* and *5ht1a* genes are expressed at significantly higher levels. Thus, different mechanisms of brain serotonergic hypofunction can be assumed in the mice with alternative social behaviors. In the losers, strong decrease of serotonergic activity may be attributed to the development of a depression-like condition with the symptoms, sensitivity to antidepressants and neurochemical changes similar to those in depressive patients [36,45,46]. In the winners, behavioral psychopathology developing under positive fighting experience [15] is associated with inhibition of brain serotonergic system. Recovering of *Tph2* and *Sert* gene expression and overexpression of the *Maoa* and *5ht1a* genes after no-fight period in the winners suggests that changes in brain serotonergic activity are not main or only cause of the behavioral pathology developing in male mice in this experimental context. Obviously, other neurotransmitter systems may also be the factors; for example, the brain dopaminergic systems, which have been found to alter in the 21-time winners. Activation of the brain dopaminergic systems has been demonstrated in male mice with prolonged positive fighting experience as elevated DOPAC (3,4-dihydroxyphenyleacetic acid) levels or/and increased DOPAC/DA (dopamine) ratios in different brain areas [15]. DA level was increased in the prefrontal cortex during and after fights in aggressive rats [8]. Upregulation of *Th* and *Dat1* gene expression has also been found in ventral tegmental area of the winners [23,47]. In the fighting deprived winners these genes retained increased expression as compared with the controls. Interestingly, in the losers' ventral tegmental area lack of significant changes in *Th* and *Dat1* expression were found [23]. Activation of *Th* and *Comt* gene expression was also found in the cerebellum, frontal cortex, hippocampus, midbrain, and striatum [48]. Thus, all data support the idea, that repeated experience of aggression results in disbalance between activities of two neurotransmitter systems - serotonergic and dopaminergic [15], namely the inhibition of the former and activation of the latter, which are part of the catecholamine systems responsible for several functions, including excitation of nervous processes. In such circumstances a low threshold for aggressive behavior has been established, and, therefore, aggression can be demonstrated even in low

provocative conditions. There is a lot of experimental evidence for the relationship between increased impulsivity and, as a consequence, increased aggression, and reduced serotonergic activity in humans and animals [9-11,49]. The results of our studies are consistent with these data.

As mentioned above, chronic positive fighting experience is accompanied by development of behavioral pathology symptoms [15,49] in male mice such as abnormal aggression, hostility, pronounced anxiety, disturbances in social recognition and motivated behavior, hyperactivity, stereotypic and hyperkinetic reactions etc. After fighting deprivation, the male mice have been shown to demonstrate higher aggressiveness as compared with a period before deprivation [20,21]. Therefore, it can be assumed that serotonergic system is not responsible for this phenomenon since serotonergic genes recover their expression after cessation of agonistic interactions. The brain dopaminergic and opioidergic systems retain their changed functional activity [15,47] and may be suspected as being the key factors of increased aggression after no-fight periods.

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